

Measurement of Intracellular Calcium Concentration in *Pseudomonas aeruginosa*

Manita Guragain, Anthony K. Campbell, and Marianna A. Patrauchan*

Department of Microbiology and Molecular Genetics, Oklahoma State University, Stillwater, US

*For correspondence: m.patrauchan@okstate.edu

[Abstract] Characterization of the molecular mechanisms of calcium (Ca^{2+}) regulation of bacterial physiology and virulence requires tools enabling measuring and monitoring the intracellular levels of free calcium ($\text{Ca}^{2+}_{\text{in}}$). Here, we describe a protocol optimized to use a recombinantly expressed Ca^{2+} -binding protein, aequorin, for detecting $\text{Ca}^{2+}_{\text{in}}$ in *Pseudomonas aeruginosa*. Upon binding to free Ca^{2+} , aequorin undergoes chromophore oxidation and emits light, the log of which intensity linearly correlates with the amount of bound Ca^{2+} , and therefore, can be used to measure the concentration of free Ca^{2+} available for binding. This protocol involves the introduction of the aequorin gene into *P. aeruginosa*, induction of apoaequorin production, reconstitution of the holoenzyme with its chromophore, and monitoring its luminescence. This protocol allows continuous measuring of $\text{Ca}^{2+}_{\text{in}}$ concentration *in vivo* in response to various stimuli.

Keywords: Intracellular calcium, Regulation, Aequorin, Luminescence, Coelenterazine, *Pseudomonas aeruginosa*

[Background] Ca^{2+} regulates physiology and virulence of *P. aeruginosa* (Guragain *et al.*, 2013; Patrauchan *et al.*, 2005; Sarkisova *et al.*, 2014), however, the molecular mechanisms of Ca^{2+} regulation are not well understood. To characterize these mechanisms, it is critically important to not only measure the concentration of $\text{Ca}^{2+}_{\text{in}}$ ($[\text{Ca}^{2+}_{\text{in}}]$), but to monitor its changes in response to various stimuli. Considering that $[\text{Ca}^{2+}_{\text{in}}]$ may change in response to even minute alterations in cell physiology (reviewed in [Dominguez *et al.*, 2015]), measuring $[\text{Ca}^{2+}_{\text{in}}]$ requires a tool specifically recognizing Ca^{2+} without significantly disturbing cells. One such tool is aequorin, a Ca^{2+} -binding protein, which upon binding to free Ca^{2+} , undergoes chromophore oxidation and emits light. The emitted light can be recorded as a measure of free Ca^{2+} . Aequorin has been successfully used to monitor $\text{Ca}^{2+}_{\text{in}}$ in eukaryotes (Bonora *et al.*, 2013), as well as several bacterial species (Herbaud *et al.*, 1998; Naseem *et al.*, 2007; Rosch *et al.*, 2008). Sufficient level of aequorin production and its stability within a cell enables continuous monitoring of $\text{Ca}^{2+}_{\text{in}}$ (Naseem *et al.*, 2007). Use of aequorin offers additional advantages such as targeted intracellular distribution (cytoplasm or periplasm), high dynamic range, high signal-to-noise ratio, and low Ca^{2+} buffering effect (Bonora *et al.*, 2013). Alternative approaches include application of chemical indicators, such as Fura. However, due to reduced cell membrane permeability in *P. aeruginosa*, loading cells of this bacterium even with membrane permeable Fura acetoxymethyl (AM, ester form) is challenging and requires additional treatments, which limits physiological relevance of the measurements (not published observations). Therefore, our group pioneered the use of aequorin for measuring $[\text{Ca}^{2+}_{\text{in}}]$ in *P. aeruginosa* (Guragain *et al.*, 2013). The original protocol was developed for

Escherichia coli (Knight *et al.*, 1991) and further developed in (Jones *et al.*, 1999). Here we present a modified adaptation of the protocol, successfully used to study Ca²⁺ homeostasis in *P. aeruginosa*, clinically and environmentally important organism (Guragain *et al.*, 2013).

Materials and Reagents

A. General supplies

1. Centrifuge bottles (No specific brand is required)
2. Microfuge tubes (No specific brand is required)
3. Lumitrac 96 well white microplates (Greiner Bio One, catalog number: 655075)
4. Aluminum foil
5. Plastic cuvettes (BrandTech Scientific, catalog number: 759086D)

B. Strains and plasmids

1. *Pseudomonas aeruginosa* strain PAO1 carrying pMBB66EH containing aequorin gene
2. Plasmid pMBB66EH encoding aequorin gene from *Aequoria victoria* (courtesy: Drs. Delfina Dominguez)

C. Culture medium

1. Luria bertani (LB) agar (see Recipes)
2. Biofilm minimal medium (BMM) (see Recipes)
 - a. 10x basal salt solution
 - b. Vitamin solution
 - c. Trace metal
 - d. 1 M MgSO₄

D. Chemicals and buffers

1. Carbenicillin, disodium (Gold Bio, catalog number: C-103-5)
2. IPTG (Gold Bio, catalog number: I2481C)
3. Calcium chloride dehydrate (CaCl₂·2H₂O) (Sigma-Aldrich, catalog number: C5080)
4. Live-Dead staining (Molecular Probes)
5. Yeast extract (BD, Bacto, catalog number: 212750)
6. Sodium chloride (NaCl) (Sigma-Aldrich, catalog number: S7653 or VWR, catalog number: E529-500ML)
7. Tryptone (BD, Bacto, catalog number: 211705)
8. Agar (BD, Bacto, catalog number: 214010)
9. Nanopure water
10. Monosodium glutamate (Sigma-Aldrich, catalogue number: 1446600)
11. Glycerol (Thermo Fisher Scientific, Fisher Scientific, catalog number: G33)

12. Sodium phosphate monobasic dihydrate (NaH_2PO_4) (Sigma-Aldrich, catalog number: 71500)
13. Potassium phosphate dibasic (K_2HPO_4) (Sigma-Aldrich, catalog number: 17835)
14. Biotin (Gold Bio, catalog number: B-950-1)
15. Thiamine HCl (Gold Bio, catalog number: T-260-25)
16. HCl (Pharmco-Aaper, catalog number: 284000ACS)
17. Copper(II) sulfate pentahydrate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) (Sigma-Aldrich, catalog number: 209198).
18. Zinc sulfate heptahydrate ($\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$) (Sigma-Aldrich, catalog number: Z0251)
19. Iron(II) sulfate heptahydrate ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$) (Sigma-Aldrich, catalog number: 215422)
20. Manganese(II) chloride tetrahydrate ($\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$) (Avantor Performance Materials, J.T. Baker, catalog number: 2540-04)
21. Magnesium sulfate heptahydrate ($\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$) (Sigma-Aldrich, catalog number: 230391)
22. HEPES (Sigma-Aldrich, catalog number: H3375)
23. MgCl_2 (Sigma-Aldrich, catalog number: 230391)
24. Tergitol Type NP-40, 70% in H_2O (Sigma-Aldrich, catalog number: T1135)
25. Coelenterazine (Thermo Fisher Scientific, Molecular Probes™, catalog number: C2944)
26. Ethanol (Pharmco-Aaper, catalog number: AAP-111000190CSGL)
27. HEPES buffer (see Recipes)
28. Discharge buffer (see Recipes)
29. 1 M CaCl_2 solution (see Recipes)
30. 6 mM CaCl_2 solution for injection (see Recipes)
31. Coelenterazine (see Recipes)

Equipment

1. Multichannel pipette (Finnipipette F1, Thermo Fisher Scientific, Thermo Scientific™, catalog number: 4661030)
2. Pipetman classic pipettes (Gilson, catalog numbers: F123600, F123615, F123601, F123602)
3. 500 ml glass flasks (No specific brand is required)
4. Synergy™ Mx multimode microplate reader (Biotek Instruments) with Gen5™ 2.05 PC software (BioTek Instruments)
5. 37 °C incubator (Bench top incubator) (VWR, catalog number: 89409-314)
6. 37 °C shaking incubator (MaxQ 4000 table top shake incubator) (Thermo Fisher Scientific, Thermo Scientific™, model: SHKA4000)
7. Centrifuge (Thermo Fisher Scientific, Thermo Scientific™, model: Sorvall™ RC 6 Plus Centrifuge) with rotor type (Thermo Fisher Scientific, Thermo Scientific™, model: F13-14x50 cy)
8. Centrifuge (Eppendorf, model: 5424)

Procedure

A. Expression and reconstitution of aequorin

1. Grow PAO1 strain carrying plasmid pMMB66EH encoding aequorin gene for overnight at 37 °C on LB agar plate with carbenicillin (300 µg/ml).
2. Inoculate 5 ml of BMM medium with 2-3 single colonies grown on LB agar plate and incubate for 12 h at 37 °C while shaking at 200 rpm. Antibiotic was omitted hereafter to avoid its effect on bacteria.
3. Transfer 1 ml of 12 h culture of OD₆₀₀ 0.25 (Adjust OD if needed by diluting in fresh BMM medium) to 100 ml of fresh BMM medium in 500 ml flask. Grow culture at 37 °C, 200 rpm until the mid-log phase, as determined in previous growth studies. For example, for PAO1, mid-log phase is reached at OD₆₀₀ of 0.16 after 12 h of incubation.
4. Once reached mid-log phase, induce cells with 1 mM IPTG (1 ml of 100 mM IPTG) and incubate for 2 h, at 37 °C, 200 rpm. At the end of induction, measure OD₆₀₀ of the culture and compare with the previously recorded growth curve, to make sure that cells are still in their logarithmic phase of growth.
5. Transfer cells into 250 ml ice cold centrifuge bottles. From this step and until the reconstitution step, maintain cells on ice.
6. Harvest cells by centrifugation at 15,000 x g for 5 min at 4 °C. Discard supernatant and wash the cell pellet with 100 ml of ice-cold HEPES buffer. For this, first carefully resuspend cells by pipetting into of 2 ml HEPES buffer, and then add the remaining 98 ml of HEPES buffer.
7. Collect cells by centrifuging at 6,000 x g for 5 min at 4 °C. Discard the supernatant.
8. Resuspend collected cell pellet in 1,250 µl of HEPES buffer by pipetting and transfer 1 ml of cell suspension into a fresh microfuge tube.
9. Add coelenterazine to a final concentration of 2.5 µM. For this, add 5 µl of 500 µM coelenterazine solution to 1 ml of cell suspension. Since coelenterazine is light sensitive, the procedure from this point onward must be carried out in the dark (see Notes).
10. After addition of coelenterazine, incubate cell suspension at room temperature without shaking for 30 min.
Note: This step is referred to as a reconstitution step. Since coelenterazine undergoes slow oxidation in the presence of atmospheric oxygen, shaking should be strictly avoided.
11. Collect the cell pellet by centrifuging at 6,000 x g for 5 min at 4 °C and wash two times with 1 ml of ice-cold HEPES buffer.
12. Resuspend the final collected cell pellet into 1 ml HEPES buffer by pipetting.
13. Measure OD₆₀₀ of thus prepared sample. Adjust the final OD₆₀₀ of the cell suspension to 0.4 by adding HEPES buffer if needed.

B. Measurement of luminescence

1. Pipette 100 μ l of cells suspension with reconstituted aequorin into 96-well luminescence (white) plate and equilibrate at room temperature for 10 min in the dark. When the effect of inhibitors or other compounds on Ca^{2+}_{in} to be tested, add the compounds during this step.
2. Load the plate with samples into a Synergy Mx plate reader, and record luminescence for 1 min, at 5 sec interval. Use this reading to calculate a basal level of Ca^{2+}_{in} .
3. To study the response of the intracellular Ca^{2+} levels to extracellular Ca^{2+} elevated to the millimolar concentrations commonly present in some environments including a human body, expose the cells prepared as above to the addition of 1 mM CaCl_2 . For this, inject 20 μ l of 6 mM CaCl_2 into each well by the plate reader injector. Prior to injection, prime the injector with 5 ml of 6 mM CaCl_2 . If the immediate effects of other compounds need to be tested, inject the latter at this step (before, after or instead of CaCl_2) through a second injector, followed by luminescence measurements.
4. Immediately after injection, mix the samples for 1 sec, and record luminescence for 20 min at 5 sec interval (Figure 1 A). Mixing and measurements were pre-programmed in the instrument.
5. In order to estimate the remaining aequorin present in the samples, briefly take out the plate from the reader, and manually add 120 μ l of discharge buffer to each sample, mix well, but quickly by pipetting, and the load back the plate into the plate reader. Read luminescence for 10 min at 5 sec interval (Figure 1B). Estimate the total aequorin by summing the luminescence detected during the entire experiment including the discharge. Use this value of total aequorin for data normalization, when calculating the concentration of Ca^{2+}_{in} (Figure 1C).

Note: it is important to ensure that no bubbles are formed during mixing, as bubbles will interfere with the luminescence reading.

Data analysis

1. Calculation of free intracellular calcium concentration ($[\text{Ca}^{2+}_{in}]$)

$[\text{Ca}^{2+}_{in}]$ (Figure 1C) was calculated from the luminescence (Figure 1A) values using the formula:
 $\text{pCa} = 0.612 (-\log_{10}k) + 3.745$

Where,

k is a rate constant for luminescence decay (sec^{-1}) as described in Jones *et al.*, 1999.

$[\text{Ca}^{2+}_{in}]$ at each time point was calculated as an average of at least three independent biological replicates. Even slight inconsistencies during harvesting and preparing cells may cause fluctuations in the $[\text{Ca}^{2+}_{in}]$ profile. Therefore, in case of inconsistent $[\text{Ca}^{2+}_{in}]$ profile, it is strongly recommended to repeat the experiment with three independent biological replicates, making sure that cells are synchronized and harvested at exactly the same point of growth. Certain mutants are particularly sensitive and produce more fluctuations in their $[\text{Ca}^{2+}_{in}]$ levels. In these cases, additional replicates were added, and a $[\text{Ca}^{2+}_{in}]$ profile, shared by at least 70% of biological replicates, was considered for further calculations. Replicates significantly deviating

from this profile were excluded. Statistical significance was calculated by standard deviations among biological replicates (Figure 1D).

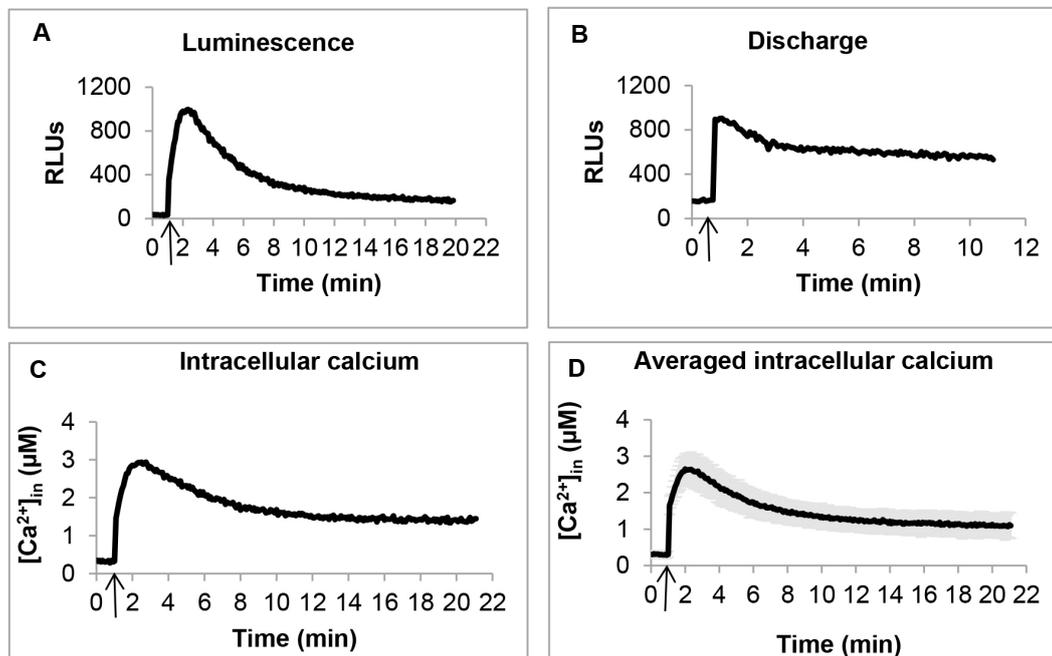


Figure 1. Measuring [Ca²⁺]_{in} in PAO1 by using aequorin. A. Representative luminescence profile. Arrow indicates addition of 1 mM CaCl₂. B. Representative discharge luminescence profile. Arrow indicates addition of discharge buffer (see Recipes). C. Representative calculated [Ca²⁺]_{in}. Arrow indicates addition of 1 mM CaCl₂. D. Averaged [Ca²⁺]_{in} calculated from three independent biological replicates. Arrow indicates addition of 1 mM CaCl₂.

Notes

- Two types of controls were included in the study
 - Control 1 was to verify that addition of HEPES buffer alone does not affect the [Ca²⁺]_{in}. For this, HEPES buffer alone was injected instead of 1 mM CaCl₂ challenge, and the entire procedure was followed as described. No buffer effect was detected.
 - Control 2 was to ensure that aequorin was not leaking through cell membranes. For this, 100 μl of cell suspension prepared for the measurements was incubated for 30 min in the dark at room temperature. Cells were removed by centrifugation 15,000 x g for 5 min, and the supernatant was collected, and mixed with 1 mM CaCl₂. Luminescence was monitored for 1 min. We did not observe any increase in luminescence during this experiment, thus confirming that there was no aequorin leakage from the cells into the supernatant. We also verified cells viability after the procedure by Live-Dead staining (Molecular Probes).

2. Luminescence profiles may vary between replicates due to possible differences in the levels of apoaequorin production. However, the calculated concentrations of $\text{Ca}^{2+}_{\text{in}}$ normalized by the total available aequorin should be consistent.
3. To ensure consistent $[\text{Ca}^{2+}]_{\text{in}}$ measurements, it is important to induce and harvest cells at the same point of growth. During our study, we followed both OD and incubation time to determine when to harvest. Only 30 min window was allowed for cells to reach the expected density, which was estimated based on prior growth analysis.
4. Aequorin is light-sensitive, and therefore all the solutions with aequorin should be kept and handled in dark tubes or tubes covered with aluminum foil. We also recommend turning off the lights in the room, only allowing either dimmed daylight or indirect light from the neighboring room. Although not tested, since aequorin absorbance maximum is at 350 nm (Shimomura and Johnson, 1969), using blue light could be safe.
5. Although aequorin is reported to be stable in solution (Prendergast, 2000), we do not recommend storing cell samples with reconstituted aequorin (even on ice) for extended period of time, as this may cause fluctuations in the luminescence profiles. For this reason, only three samples were monitored at a time.

Recipes

1. Luria Bertani (LB) agar
 - 5 g yeast extract
 - 5 g NaCl
 - 10 g tryptone
 - 15 g agarCombine the ingredients in 1 L of nanopure water and autoclave
2. Biofilm minimal media (BMM) (Patrauchan *et al.*, 2005)
 - Mix 100 ml of sterile 10x basal salt solution (Recipe 2a) to 900 ml of sterile nanopure water. Add 1 ml of vitamin solution (Recipe 2b), 200 μl of trace metals solution (Recipe 2c), and 20 μl of MgSO_4 (Recipe 2d). Mix properly.
 - a. 10x basal salt solution (9.0 mM sodium glutamate, 50 mM glycerol, 0.15 mM NaH_2PO_4 , 0.34 mM K_2HPO_4 , 145 mM NaCl):
 - 15 g monosodium glutamate
 - 46 g glycerol
 - 0.18 g sodium phosphate monobasic dehydrate (NaH_2PO_4)
 - 0.78 g potassium phosphate dibasic (K_2HPO_4)
 - 84.7 g sodium chloride (NaCl)Combine the ingredients and dissolve completely in 850 ml nanopure water. Adjust the pH to 7.0. Adjust the final volume to 1,000 ml with nanopure water. Sterilize by autoclaving.
 - b. Vitamin solution

Dissolve 1 mg of biotin in 10 ml of nanopure water. Aliquot 1 ml of biotin stock solution in fresh tube and add 50 mg thiamine HCl to it and mix properly. Adjust the final volume to 100 ml with nanopure water. Filter sterilize and store at 4 °C.

c. Trace metal

Dilute 10 ml concentrated HCl into 70 ml of nanopure water. Add the following ingredients:

0.5 g CuSO₄ 5H₂O

0.5 g ZnSO₄ 7 H₂O

0.5 g FeSO₄ 7H₂O

0.2 g MnCl₂ 4H₂O

Dissolve completely and adjust the final volume to 100 ml with nanopure water. Filter-sterilize or autoclave.

d. 1 M MgSO₄

24.64 g of MgSO₄·7H₂O was dissolved in final volume of 100 ml of nanopure water. The solution was sterilized by autoclaving.

3. HEPES buffer (25 mM HEPES, 125 mM NaCl, 1 mM MgCl₂, pH7.5)

5.96 g HEPES

7.3 g NaCl

0.0952 MgCl₂

Dissolve the ingredients in 900 ml nanopure water. Adjust the pH to 7.5 and titrate to the final volume of 1,000 ml with nanopure water.

4. Discharge buffer (12.5 mM CaCl₂, 2% NP-40 in HEPES buffer)

To 5 ml of HEPES buffer, add 62.5 µl of 1 M CaCl₂ solution and 143 µl of Tergitol. Gently mix by stirring on a magnetic stir plate to avoid foaming of NP-40 detergent

5. 1 M CaCl₂ solution

Dissolve 36.75 g of CaCl₂ dihydrate in 250 ml nanopure water. Sterilize by autoclaving

6. 6 mM CaCl₂ solution for injection

240 µl of 1 M CaCl₂ solution was added to 39.76 ml of HEPES buffer and mixed properly

7. Coelenterazine

Pulse-centrifuge the tube to collect the entire quantity of the reagent (250 µg coelenterazine) at the bottom. Add 1,136 µl of 95% ethanol to the tube and mix by pipetting thoroughly but quickly (to avoid light and ethanol evaporation). Resting the tube (closed) on bench for a few minutes helps dissolving. After dissolving, aliquot 50 µl of the coelenterazine solution into 0.5 ml microfuge tubes. Cover the microfuge tubes with aluminum foil and store at -20 °C

Note: Coelenterazine is highly light sensitive and should be handled in the dark.

Acknowledgments

We thank Dr. Delfina Dominguez from The University of Texas at El Paso for sharing *E. coli* strain carrying pMMB66EH. We thank Ian Reutlinger for transformation of *P. aeruginosa* PAO1 strains with

pMMB66EH plasmid containing aequorin gene. This work was supported by the Grant-in-Aid from American Heart Association (Award 09BGIA2330036) and the Research Grant from OCAST (Award HR12-167).

References

1. Bonora, M., Giorgi, C., Bononi, A., Marchi, S., Patergnani, S., Rimessi, A., Rizzuto, R. and Pinton, P. (2013). [Subcellular calcium measurements in mammalian cells using jellyfish photoprotein aequorin-based probes](#). *Nat Protoc* 8(11): 2105-2118.
2. Dominguez, D. C., Guragain, M. and Patrauchan, M. (2015). [Calcium binding proteins and calcium signaling in prokaryotes](#). *Cell Calcium* 57(3): 151-165.
3. Guragain, M., Lenaburg, D. L., Moore, F. S., Reutlinger, I. and Patrauchan, M. A. (2013). [Calcium homeostasis in *Pseudomonas aeruginosa* requires multiple transporters and modulates swarming motility](#). *Cell Calcium* 54(5): 350-361.
4. Herbaud, M. L., Guiseppi, A., Denizot, F., Haiech, J. and Kilhoffer, M. C. (1998). [Calcium signalling in *Bacillus subtilis*](#). *Biochim Biophys Acta* 1448(2): 212-226.
5. Jones, H. E., Holland, I. B., Baker, H. L. and Campbell, A. K. (1999). [Slow changes in cytosolic free Ca²⁺ in *Escherichia coli* highlight two putative influx mechanisms in response to changes in extracellular calcium](#). *Cell Calcium* 25(3): 265-274.
6. Knight, M. R., Campbell, A. K., Smith, S. M. and Trewavas, A. J. (1991). [Recombinant aequorin as a probe for cytosolic free Ca²⁺ in *Escherichia coli*](#). *FEBS Lett* 282(2): 405-408.
7. Naseem, R., Davies, S. R., Jones, H., Wann, K. T., Holland, I. B. and Campbell, A. K. (2007). [Cytosolic Ca²⁺ regulates protein expression in *E. coli* through release from inclusion bodies](#). *Biochem Biophys Res Commun* 360(1): 33-39.
8. Patrauchan, M. A., Sarkisova, S., Sauer, K. and Franklin, M. J. (2005). [Calcium influences cellular and extracellular product formation during biofilm-associated growth of a marine *Pseudoalteromonas* sp.](#) *Microbiology* 151(Pt 9): 2885-2897.
9. Prendergast, F. G. (2000). [Structural biology: Bioluminescence illuminated](#). *Nature* 405(6784): 291-293.
10. Rosch, J. W., Sublett, J., Gao, G., Wang, Y. D. and Tuomanen, E. I. (2008). [Calcium efflux is essential for bacterial survival in the eukaryotic host](#). *Mol Microbiol* 70(2): 435-444.
11. Sarkisova, S. A., Lotlikar, S. R., Guragain, M., Kubat, R., Cloud, J., Franklin, M. J. and Patrauchan, M. A. (2014). [A *Pseudomonas aeruginosa* EF-hand protein, EfhP \(PA4107\), modulates stress responses and virulence at high calcium concentration](#). *PLoS One* 9(2): e98985.
12. Shimomura, O. and Johnson, F. H. (1969). [Properties of the bioluminescent protein aequorin](#). *Biochemistry* 8(10): 3991-3997.