

Semi-denaturing Detergent Agarose Gel Electrophoresis (SDD-AGE)

Laura Molina García¹ and Fátima Gasset-Rosa^{2*}

¹Department of Cellular and Molecular Biology, Centro de Investigaciones Biológicas-CSIC, Madrid, Spain; ²Department of Neurosciences, University of California, San Diego, USA

*For correspondence: fgasset@ucsd.edu

[Abstract] Pathological proteins in neurodegenerative diseases suffer a conformational change to a misfolded amyloid state. Such pathological event leads to the aggregation of these proteins that indefinitely propagates as an altered form of itself, and harbor prion-like properties (Wickner, 1994; Prusiner, 2012). In addition to diseases, prions can also have beneficial adaptive roles in lower eukaryotes (in fungi and yeast) (Eaglestone *et al.*, 1999; True *et al.*, 2004; Coustou *et al.*, 1999). Besides separating polymers from their precursor soluble monomers, another particular difficulty of the study of amyloid proteins is to resolve the heterogeneity of the aggregates, since these usually exhibit a variable degree of polymorphism. Semi-denaturing detergent agarose gel electrophoresis (SDD-AGE) is a technique that takes advantage of both the property of prions and prion-like polymers to be highly resistant to solubilization by SDS detergent, and the large pores sizes of agarose, that allow the resolution of high molecular weight complexes. In this method, we describe in detail how this technique can be used to characterize heterogeneous aggregation in bacteria and yeast (Gasset-Rosa *et al.*, 2014; Molina-García and Giraldo, 2014), and further be applied to study the aggregation pattern of proteins that become prone to aggregation through genetic manipulation.

Materials and Reagents

1. Bacteria: Gram negative *Escherichia coli* (*E. coli*) or yeast *Saccharomyces cerevisiae* (*S. cerevisiae*)

Notes:

Amyloids exist in E. coli and S. cerevisiae. In both organism amyloids can be functional. In E. coli the most well know example is the curli protein which is assembled in the extracellular medium and allow the attachment of the bacteria to the surface and sustain the formation of biofilms (Chiti and Dobson, 2006; Chapman et al., 2002).

In S. cerevisiae, amyloids confer to cells selective advantage under certain physiological conditions and they aren't harmful (Chien et al., 2004; Shorter and Lindquist, 2005; Wickner et al., 2007), a few examples are Sup35p y Ure2p, Rnq1p. The conversion of Sup35 from soluble to the amyloid state drives the reduction of translation termination

- activity (Wickner et al., 1995).*
2. Protease inhibitor cocktail (Roche Diagnostics, catalog number: 04 693 159 001)
 3. Silica beads (1 mm lysing matrix C) (MP Biomedicals, catalog number: 6912-050)
 4. Pure agarose D2 (Pronadisa, catalog number: 8034)
 5. Detergents: SDS (Bio-Rad Laboratories, catalog number: 161-0301) and Sarkosyl 97% N-lauroylsarcosine (Sigma-Aldrich, catalog number: L-9150)
 6. PVDF membrane (Bio-Rad Laboratories)
 7. Antibodies

Notes:

Depends on the protein you work, the antibody may be against the protein that you are studying for aggregation. A recommended strategy if the user is going to generate a recombinant protein is to include a tag in the amino or carboxyl terminal of the protein and use a specific antibody against the tag. One example is the Histidine-tag, the corresponding antibody would be a Monoclonal anti-polyhistidine (Sigma-Aldrich, catalog number: H1029).

Antibodies that recognize monomer state may still be able to identify the protein in polymers even is the protein conformation has changed. Usually monomer antibodies have a broad range recognition epitopes. In our hands the antibodies used were:

For E. coli, house-made polyclonal antibodies against RepA (generated in rabbit immunized with soluble RepA), (Gasset-Rosa et al., 2014); and monoclonal anti poly-histidine against His-RepA (WH1) (Sigma-Aldrich, catalog number: H1029) (Molina-García and Giraldo, 2014). Both of them work.

For S. cerevisiae, anti-HA antibody (Roche Diagnostics, catalog number: 12CA5) against the HA ([Human influenza hemagglutinin](#)) tag of Sup35 protein (unpublished).

8. Cell culture medium
 - a. Luria Bertani (LB) medium (see Recipes)
 - b. Yeast extract peptone dextrose (YPD) (see Recipes)
9. Lysis buffer (see Recipes)
10. Running buffer (see Recipes)
11. Loading buffer (see Recipes)
12. Transference buffer (see Recipes)

Equipment

1. Fast Prep-MilliPore/MP FastPrep-24 homogenizer (catalog number: 6004-500)
2. Horizontal agarose electrophoresis system (Electrophoresis power supply EPS601 General Electrics)

3. Wet/Tank transfer blotting system (Bio-Rad Laboratories)

Procedure

1. Growth cells in your adjusted conditions.
2. Harvest cells.
 - a. Bacteria: Harvest cells (25 ml O.D. = 2) by centrifugation (10 min, 3,500 rpm, 4 °C). And, resuspend the cells in 400 µl of lysis buffer.
 - b. Yeast: Harvest cells (200 ml O.D. = 0.8) by centrifugation (5 min, 3,500 rpm, 4 °C). And, resuspend the cells in 500 µl of lysis buffer.
3. Transfer the suspension to a tube containing silica beads (usually 1/3 of the volume is sufficient).
4. Lysate the cells.
 - a. Bacteria: Lysate the cells using the Fast-Prep 24 homogenizer (4 cycles at speed IV, 30 sec each at 4 °C). And, dilute 3 µl of cell lysate with 30 µl with lysis buffer, and then add 10 µl of 4x loading buffer to the sample.
 - b. Yeast: Lysate the cells using the Fast-Prep 24 homogenizer (5 cycles at speed V, 30 sec each at 4 °C). And, add 10 µl of 4x loading buffer to 30 µl of the lysate.
5. Incubate the samples 10 min at room temperature.
6. Load them in the agarose gel.
 - a. Prepare 200 ml of agarose at 1.5% in 1x TAE.
Note: Heat the agarose suspension to melting and then add SDS to 0.1%.
 - b. Pour it into 20 cm x 24 cm horizontal plate mould. Make sure that the casting tray is on a flat surface.
Notes:
 - i. *Eliminate all the possible bubbles.*
 - ii. *Agarose electrophoresis are run using horizontal agarose electrophoresis system, the samples need to be run for long distance in order to resolve polymers of high molecular weight.*
 - c. Fit a thick comb (e.g., >2 mm, 20-well) allowing to load 40 µl per sample.
 - d. Submerge the gel in a horizontal electrophoresis container with 1x TAE-0.1% SDS at 10 °C.
7. Run electrophoresis at 100 V for 7.5 h at 10 °C.
Note: For detecting oligomers with high hydrophobic feature run it at lower voltage 50 V and room temperature for 12 h with 1% (final concentration) of Sarkosyl in the sample (4% Sarkosyl in the loading buffer). The concentration for the running buffer is TAE 1x and 0.1% SDS as described before.

Caution: Extreme care during electrophoresis to avoid any electrical hazard.

8. Cut the PVDF membrane adjusted to the size of the agarose gel and 2 pieces with the same dimensions of Whatman paper.
9. Attach tightly the gel to the PDVF membrane and add in each side a piece of Whatman paper. *Note: Everything should be pre-wet with transference buffer. Make sure that there are no bubbles in between.*
10. Place the cast into the Wet Transfer-Blot and cover with buffer 1x TAE-0.1% SDS.

Caution: Extreme care during electrophoresis to avoid any electrical hazard.

11. Run the transference at 16 V/400 mA and 10 °C for 15 h.
12. Perform Western-blot with the appropriate antibodies to detect your polymers.

Recipes

1. Luria Bertani (LB) medium
 - Bacto-tryptone 10 g/L
 - Yeast extract 5 g/L
 - NaCl 5 g/L
2. Yeast extract peptone dextrose (YPD)
 - 2% bacto peptone
 - 2% glucose
 - 1% yeast extract
3. Lysis buffer
 - 25 mM Tris-HCl (pH 6.8)
 - 250 mM NaCl
 - 5 mM EDTA
 - 10% glycerol supplemented with protease inhibitors (1 tablet each 10 ml)
4. Running buffer
 - 1x TAE (Tris acetate EDTA)
 - 0.1% SDS
5. Loading buffer (4x)
 - 0.5% TAE
 - 5% glycerol
 - 2% sarkosyl
 - 0.1 mg/ml bromophenol blue plus protease inhibitors (1 tablet each 10 ml)
6. Transference buffer
 - 1x TAE (Tris Acetate EDTA)
 - 0.1% SDS

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