

Measurement of Endogenous MALT1 Activity

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[Abstract] MALT1 (Mucosa associated lymphoid tissue protein 1) is an important adapter protein for the NF- κ B driven lymphocyte activation and the development and survival of distinct B-cell lymphoma entities. In addition MALT1 is a cysteine protease that structurally resembles caspases while having a different substrate preference and mechanism of activation. This paracaspase activity of MALT1 has been shown to be critical for an optimal NF- κ B activation and survival of the aggressive ABC-DLBCL (activated B cell-type of diffuse large B cell lymphoma), which highlights the protease as an attractive therapeutic target for the treatment of distinct B-cell lymphomas and immune diseases like rheumatoid arthritis or multiple sclerosis. In this protocol we describe a fluorogenic cleavage assay, which can be used to measure endogenous and also ectopic MALT1 activity. To this end, cellular MALT1 needs to be precipitated from the lysed cells via antibody immunoprecipitation and subsequently incubated with a fluorogenic substrate peptide. The MALT1 cleavage assay has been developed to directly determine the activity profile of MALT1 in the course of the adaptive immune response as well as in pathological signaling in lymphoid malignancies. In addition, the MALT1 activity assay has been successfully used to monitor cellular MALT1 inhibition with small molecule inhibitors.

Materials and Reagents

1. Primary human and murine cells
2. T-cell-lines (Jurkat)
3. B-cell-lines: ABC-(TMD8, HBL1, OCI-Ly3, U2932, OCI-Ly10, RIVA) and GCB-DLBCL (Su-DHL-6, Su-DHL-4, BJAB)
4. RPMI 1640 (Life Technologies, catalog number: 21875)
5. IMDM (Life Technologies, catalog number: 21056)
6. Fetal Bovine Serum (FBS) (Life Technologies, catalog number: 10270)
7. PMA (Phorbol 12-Myristat 13-Acetate) (Calbiochem, catalog number: 16561-29-8)
8. Lonomycin (Calbiochem, catalog number: 407950)
9. Anti-CD3/CD28 (Hit3a/CD28.2) (BD Heidelberg)
10. IgG1 (R19-15) (BD Pharmingen, catalog number: 553440)
11. IgG2a (A85-1) (BD Pharmingen, catalog number: 553387)

12. MALT1 antibody (Santa Cruz, catalog number: H300)
13. Protein-G Sepharose (GE Healthcare, catalog number: 17-0618-01)
14. PBS (Life Technologies, catalog number: 14190)
15. Ac-LRSR-AMC (Peptides International, catalog number: MCA-3952-PI)
16. Z-VRPR-FMK (ENZO Life Sciences, catalog number: ALX-260-166)
17. Complete protease inhibitor cocktail tablets (Roche, catalog number: 11836145001)
18. Mepazine (Hit2lead, catalog number: 5216177)
19. 384-well non-binding plates, black (Greiner Bio-one, catalog number: 781900)
20. Saccharose
21. CHAPS
22. HEPES
23. Triton X-100
24. Cellular lysis buffer (see Recipes)
25. MALT1 cleavage buffer (see Recipes)

Equipment

1. 37 °C 5% CO₂ cell culture incubator
2. 26 G syringe (Roth, catalog number: C718.1)
3. Centrifuge (Eppendorf, model: 5417R)
4. Synergy II multiwell plate reader (Biotek)
5. Rotary Mixer

Procedure

1. To activate MALT1 plate 2.5×10^6 cells (Jurkat T-cells, primary T cells or PBMCs) and either stimulate with PMA/Ionomycin (200 ng/ml and 300 ng/ml final concentrations) in 10 ml of complete RPMI media in 25 cm² cell culture flasks or with anti-CD3 and anti-CD28 antibodies (1 µg/ml final concentration each) in the presence of anti-IgG1/IgG2a coupling antibodies (0.5 µg/ml each) in 1 ml of complete RPMI media in 12-well for 30 min or treat with solvent (H₂O) in the control. Cells with constitutive MALT1 activity (e.g. ABC-DLBCL) can be left untreated.
2. As a negative control treat the above cells with MALT1 inhibitors, e.g. the tetrapeptide Z-VRPR-FMK (50 µM) or small molecule inhibitors like mepazine (10 µM) 3 - 6 h prior to stimulation and lysis.
3. All 2.5×10^6 cells per reaction are then pelleted at 300 x g for 5 min.

4. Removal of supernatant and lysis of the cells with cellular lysis buffer (500 μ l) by rotating for 20 min on 4 °C in a 1.5 ml reagent tube.
5. Centrifugation of the lysates for 10 min at 21,000 x g to remove cell debris.
6. Incubation of all of the lysate with 700 ng MALT1 antibody over night at 4 °C on a rotary mixer.
7. Incubation with 15 μ l Protein-G sepharose beads (beads were washed and diluted 1:2 in PBS and equilibrated before usage) for 60 min at 4 °C on a rotary mixer.
8. Beads are pelleted and washed 3 times with PBS at 300 x g for 2 min at 4 °C.
9. In the last washing step all PBS is discarded via a syringe and 45 μ l cleavage buffer is added to the beads.
10. The beads are pelleted at 300 x g for 1 min afterwards re-suspended carefully and 49 μ l of the suspension (beads and cleavage buffer) are subsequently transferred to one well of a 384-well assay plate.
11. 1 μ l of the fluorogenic substrate Ac-LRSR-AMC is then added to the wells in a final concentration of 20 μ M and the plates are placed into the plate reader where they are shaken for 10 sec to mix the reagents.
12. After an incubation of 30 min at 30 °C the release of AMC fluorescence due to MALT1 cleavage is measured at 360 nm excitation and 460 nm emission over a time course of 90 min.
13. Specificity of MALT1 cleavage activity *in vitro* can be assessed by adding 5 nM of Z-VRPR-FMK to a separate control reaction (negative control).

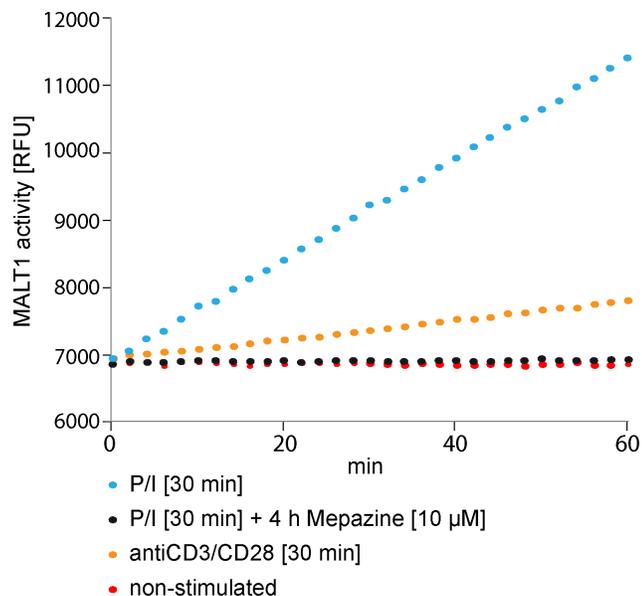


Figure 1. Fluorogenic MALT1 cleavage assay. Jurkat T-cells (2.5×10^6) were pre-incubated with Mepazine or DMSO for 4 h and subsequently stimulated with

PMA/Ionomycin (P/I) or antiCD3/CD28 for 30 min or left untreated. After lysis of the cells MALT1 was precipitated and the catalytic activity was measured after addition of Ac-LRSR-AMC in a Synergy II plate reader over 60 min at 2 min intervals.

Recipes

1. Cellular lysis buffer
 - 50 mM HEPES (pH 7.5)
 - 10% (v/v) Glycerin
 - 0.1% (v/v) Triton X-100
 - 1 mM Dithiothreitol (DTT)
 - 150 mM NaCl
 - 2 mM MgCl₂
 - 1 complete protease inhibitor tablet per 50 ml
2. MALT1 cleavage buffer
 - 50 mM MES (pH 7.0)
 - 150 mM NaCl
 - 10% (w/v) Saccharose
 - 0.1% (w/v) CHAPS
 - 1 M Sodium citrate
 - 10 mM DTT

Acknowledgments

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References

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