

Structural Based Strategy for Predicting Transcription Factor Binding Sites

Beisi Xu^{1*}, Yongmei Wang², Haojun Liang³ and Guohui Li³

¹State Key Laboratory of Molecular Reaction Dynamics, Dalian Institute of Chemical Physics, The Chinese Academy of Sciences, Dalian, Liaoning, China; ²Department of Chemistry, University of Memphis, Memphis, TN, USA; ³Department of Polymer Science and Engineering, University of Science and Technology of China, Hefei, Anhui, China

*For correspondence: xubeisi@gmail.com

[Abstract] Scanning through genomes for potential transcription factor binding sites (TFBSs) is becoming increasingly important in this post-genomic era. The position weight matrix (PWM) is the standard representation of TFBSs utilized when scanning through sequences for potential binding sites. Many transcription factor (TF) motifs are short and highly degenerate, and methods utilizing PWMs to scan for sites are plagued by false positives. Furthermore, many important TFs do not have well-characterized PWMs, making identification of potential binding sites even more difficult. One approach to the identification of sites for these TFs has been to use the 3D structure of the TF to predict the DNA structure around the TF and then to generate a PWM from the predicted 3D complex structure. However, this approach is dependent on the similarity of the predicted structure to the native structure. We introduce here a novel approach to identify TFBSs utilizing structure information that can be applied to TFs without characterized PWMs, as long as a 3D complex structure (TF/DNA) exists. Our approach utilizes an energy function that is uniquely trained on each structure thus leads to increased prediction accuracy and robustness compared with those using a more general energy function. The software is freely available upon request. Please see reference supplementary material for details.

Data and Software

A. TF/DNA structure

tFIRE takes standard PDB format for TF/DNA structures.

1. A good place to look for such data is PDB (Rose *et al.*, 2011) (<http://www.pdb.org>).
2. If the TF/DNA complex structure does not exist but the TF structure exists, you can generate the TF/DNA structure by docking DNA to the TF structure. Software that can be utilized for this includes HADDOCK (De Vries *et al.*, 2007) (<http://www.nmr.chem.uu.nl/haddock>), FTDOCK (Jackson R.M. *et al.*, 1998) (<http://www.sbg.bio.ic.ac.uk/docking/ftdock.html>), YASARA DOCK (<http://www.yasara.org/dnadock.htm>), ParaDock (Banitt and Wolfson, 2011)

(<http://www.paradocks.org>). Our method indicates that such docking will not affect a result significantly but we have not tested any of these docking predictions ourselves for validation. Hence we urge their use with caution, and revalidate the results once the structures are available.

3. If the TF structure does not exist in 3D structure databases, you can predict TF structure using homology modeling like SWISS-MODEL (Guex and Peitsch, 1997) (<http://swissmodel.expasy.org>), Rosetta (Bradley *et al.*, 2003) (<https://www.rosettacommons.org>), Sybyl (Visegrády *et al.*, 2001) (<http://www.tripos.com/index.php?family=modules,SimplePage,,,&page=SYBYL-X>).

Please use with caution that each prediction step would reduce the accuracy.

B. Predict TFBSs

tFIRE predicted motif(PWM) can be used to predict TFBSs.

1. Motif scanning programs can be used to scan the whole genome for motif matches. Such methods included MAST (Bailey *et al.*, 2006) (<http://meme.nbcr.net/meme/cgi-bin/mast.cgi>) from MEME suite and STORM (Schones *et al.*, 2007) (<http://rulai.cshl.edu/storm>) from Cold Spring Harbor Laboratory.
2. TFBSs vary for different cells. Recently, a newly developed method, CENTIPEDE (Pique-Regi *et al.*, 2010) (<http://centipede.uchicago.edu>) shows that with the result from a single DNase-seq experiment, one can accurately predict TFBSs for all TFs. Therefore, downloading DNase-seq data from ENCODE project can be very helpful.
3. Recently developed FAIRE-seq technology allow similar predictions for detection of chromatin accessibility regions (Song *et al.*, 2011). Such data can be substituted for DNase-seq, but needs be tested and validated before use.
4. Epigenetic information can also be employed instead of DNase-seq (Cuellar-Partida *et al.*, 2012). We propose to update our data and methods available for such predictions on a regular basis.

C. Software

1. C++ software environment, better with Linux system.
2. tFIRE, Feel Free to ask the author for a linux version.
3. WebLogo (Crooks *et al.*, 2004) can be used for visualization the PWM we predicted. (<http://weblogo.berkeley.edu>)

Procedure

1. If you are confident of your TF/DNA complex 3D structure, then you can use tFIRE default function pre-trained by all available TF/DNA structures in the PDB database. You can also construct your own energy function with tFIRE by several non-homology structures. You can use the PISCES server (Wang and Dunbrack, 2003) (<http://dunbrack.fccc.edu/PISCES.php>) that this server will give you a subset of your input structure list (PDB id) that each protein in the subset has little homology to another.
2. If you are not confident of your TF/DNA structure, you can train tFIRE with a single structure and subsequently predict PWMs using tFIRE.

Acknowledgments

This protocol has been adapted from: Xu *et al.* (2013). We thank the funding supported by the National Sciences Foundation of China (no. 31070641) and National 973 Program of China (no. 2012CB721000) and start-up funding from SKLMRD and DICP, CAS (Chinese Academy of Sciences). The funders offered most of the costs of study design, data collection and analysis, decision to publish, or preparation of the manuscript. We would like to thank Dr. Yan Cui, Dr. Yaoqi Zhou, Dr. Yuedong Yang, Dr. Chi Zhang, Dr. Song Liu, Dr. Jason Donald, Dr. Eugene Shakhnovich, Dr. Timothy Robertson, Dr. Gabriele Varani, Dr. Marc Jung, Dr. Amy Leung and Dr. Rongze Lu, Juan Du for their databases, programs and helpful discussions.

References

1. Bailey, T. L., Williams, N., Misleh, C. and Li, W. W. (2006). [MEME: discovering and analyzing DNA and protein sequence motifs](#). *Nucleic Acids Res* 34(Web Server issue): W369-373.
2. Banitt, I. and Wolfson, H. J. (2011). [ParaDock: a flexible non-specific DNA-rigid protein docking algorithm](#). *Nucleic Acids Res* 39(20): e135.
3. Bradley, P., Chivian, D., Meiler, J., Misura, K. M., Rohl, C. A., Schief, W. R., Wedemeyer, W. J., Schueler-Furman, O., Murphy, P., Schonbrun, J., Strauss, C. E. and Baker, D. (2003). [Rosetta predictions in CASP5: successes, failures, and prospects for complete automation](#). *Proteins* 53 Suppl 6: 457-468.
4. Crooks, G. E., Hon, G., Chandonia, J. M. and Brenner, S. E. (2004). [WebLogo: a sequence logo generator](#). *Genome Res* 14(6): 1188-1190.

5. Cuellar-Partida, G., Buske, F. A., McLeay, R. C., Whittington, T., Noble, W. S. and Bailey, T. L. (2012). [Epigenetic priors for identifying active transcription factor binding sites](#). *Bioinformatics* 28(1): 56-62.
6. Guex, N. and Peitsch, M. C. (1997). [SWISS-MODEL and the Swiss-PdbViewer: an environment for comparative protein modeling](#). *Electrophoresis* 18(15): 2714-2723.
7. Jackson, R. M., Gabb, H. A. and Sternberg, M. J. (1998). [Rapid refinement of protein interfaces incorporating solvation: application to the docking problem](#). *J Mol Biol* 276(1): 265-285.
8. Pique-Regi, R., Degner, J. F., Pai, A. A., Gaffney, D. J., Gilad, Y. and Pritchard, J. K. (2011). [Accurate inference of transcription factor binding from DNA sequence and chromatin accessibility data](#). *Genome Res* 21(3): 447-455.
9. Rose, P. W., Beran, B., Bi, C., Bluhm, W. F., Dimitropoulos, D., Goodsell, D. S., Prlic, A., Quesada, M., Quinn, G. B., Westbrook, J. D., Young, J., Yukich, B., Zardecki, C., Berman, H. M. and Bourne, P. E. (2011). [The RCSB Protein Data Bank: redesigned web site and web services](#). *Nucleic Acids Res* 39(Database issue): D392-401.
10. Schones, D. E., Smith, A. D. and Zhang, M. Q. (2007). [Statistical significance of cis-regulatory modules](#). *BMC Bioinformatics* 8: 19.
11. Song, L., Zhang, Z., Grasfeder, L. L., Boyle, A. P., Giresi, P. G., Lee, B. K., Sheffield, N. C., Graf, S., Huss, M., Keefe, D., Liu, Z., London, D., McDaniell, R. M., Shibata, Y., Showers, K. A., Simon, J. M., Vales, T., Wang, T., Winter, D., Clarke, N. D., Birney, E., Iyer, V. R., Crawford, G. E., Lieb, J. D. and Furey, T. S. (2011). [Open chromatin defined by DNaseI and FAIRE identifies regulatory elements that shape cell-type identity](#). *Genome Res* 21(10): 1757-1767.
12. Visegrady, B., Than, N. G., Kilar, F., Sumegi, B., Than, G. N. and Bohn, H. (2001). [Homology modelling and molecular dynamics studies of human placental tissue protein 13 \(galectin-13\)](#). *Protein Eng* 14(11): 875-880.
13. de Vries, S. J., van Dijk, A. D., Krzeminski, M., van Dijk, M., Thureau, A., Hsu, V., Wassenaar, T. and Bonvin, A. M. (2007). [HADDOCK versus HADDOCK: new features and performance of HADDOCK2.0 on the CAPRI targets](#). *Proteins* 69(4): 726-733.
14. Wang, G. and Dunbrack, R. L., Jr. (2003). [PISCES: a protein sequence culling server](#). *Bioinformatics* 19(12): 1589-1591.
15. Xu, B., Yang, Y., Liang, H. and Zhou, Y. (2009). [An all-atom knowledge-based energy function for protein-DNA threading, docking decoy discrimination, and prediction of transcription-factor binding profiles](#). *Proteins* 76(3): 718-730.
16. Xu, B., Schones, D. E., Wang, Y., Liang, H. and Li, G. (2013). [A structural-based strategy for recognition of transcription factor binding sites](#). *PLoS One* 8(1): e52460.