

LC3B Labeling on Terrestrial Isopod Adipocytes

Christine Braquart-Varnier^{*}, Maryline Raimond and Mathieu Sicard

Ecologie et Biologie des Interactions, UMR 7267 CNRS, Université de Poitiers, Poitiers Cedex, France

*For correspondence: christine.braquart@univ-poitiers.fr

[Abstract] The LC3B protein plays a critical role in autophagy. Normally, this protein resides in the cytosol, but following cleavage and lipidation with phosphatidylethanolamine, LC3B associates with the phagophore. This localization can be used as a general marker for autophagic membranes. To visualize the LC3B, we used the LC3B Antibody Kit for Autophagy (Invitrogen). As this kit was designed to work with cell coming from cell culture it was not possible to perform it on compact tissues such as nerve cord or ovaries. We thus adapted LC3B labeling initial protocol to be able to label autophagic membranes of adipocytes from terrestrial isopods which form a loose tissue that can be more easily penetrated by antibodies. The following protocol permits a visualization of the autophagic vesicles directly in terrestrial isopod cells freshly sampled in animals.

Materials and Reagents

1. LC3B Antibody Kit for Autophagy (Life Technologies, Invitrogen™, catalog number: L10382)
2. Alexa fluor 555 goat anti-rabbit IgG (H + L) 2 mg/ml (Life Technologies, Invitrogen™, catalog number: A21428)
3. Formaldehyde
4. BSA
5. DAPI (2.5 µg/ml) (Sigma-Aldrich)
6. AF1 antifading-25 ml (Citifluor, England)
7. Pincers for dissection
8. Pap Pen Liquid Blocker Super Pap Pen (Daido Sangyo Co. Ltd, Japan)
9. Glass slide
10. Blocking buffer (see Recipes)
11. Terrestrial isopod Phosphate buffered saline (PBS) (see Recipes)
12. Fixative (see Recipes)
13. Permeabilization reagent (see Recipes)

Equipment

1. Black box
2. Epifluorescent microscope (Axio Observer-A1, Zeiss) with Apotome (structured illumination) equipped with a 63x/1.25 objective (oil immersion) and with the AxioVision 4.8.1 software (Zeiss)

Procedure

1. Dilute the LC3B rabbit polyclonal antibody in blocking buffer to prepare 0.5 µg/ml working solution (1 µl stock solution in 1,999 µl of blocking buffer).
2. Draw a well (of a diameter of around 1 cm) with a liquid repellent pen on glass slide.
3. Dissect the terrestrial isopod and collect fat tissue especially those surrounding the nerve cord (a length of around 6 mm) (Figure 1).

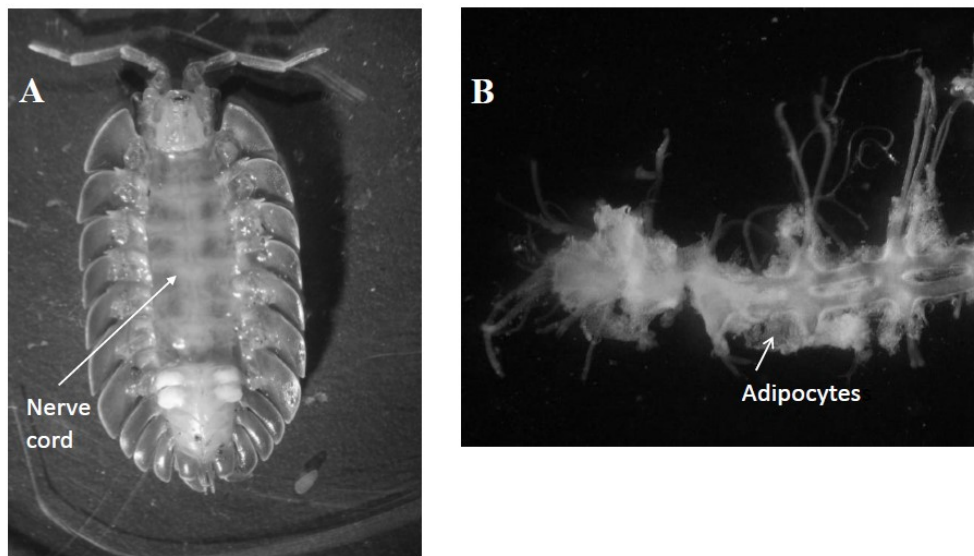


Figure 1. Location of the nerve cord in *Porcellio dilatatus* (A) and the nerve cord surrounded by adipocytes (Ad) once collected (B).

4. Deposit the nerve cord surrounded by adipocytes (fat tissue) on the middle of the drawn well.
5. Add 150 µl of the 3.7% formaldehyde fixative in terrestrial isopod PBS onto the tissue.
6. Incubate 15 min in a black box containing a humid paper at room temperature (RT).
7. Remove the fixative and wash the tissue three times with terrestrial isopod PBS (5 min with 150 µl of PBS by washing).

8. Remove the PBS after the last wash.
9. Prepare 0.2% Triton X-100 in terrestrial isopod PBS and apply 150 onto the fat tissue.
10. Be careful of not dissolving the liquid repellent pen for the tissue to be always immersed in the solution.
11. Incubate in a black box with humid paper at 15 min RT.
12. Draw a well (diameter, ~1 cm) with a liquid repellent pen on a fresh glass slide.
13. Collect the fat tissue with forceps and transfer it onto the fresh slide.
14. Add 150 μ l of the diluted primary antibody prepared in step 1 onto the fat tissue.
15. Incubate 60 min in a black box containing a humid paper at RT.
16. Remove the primary antibody solution and wash the tissue three times with terrestrial isopod PBS (5 min with 150 μ l of PBS by washing).
17. Remove the PBS after the last washing.
18. Dilute the goat anti-rabbit secondary antibody in blocking buffer to prepare 5 μ g/ml working solution (1 μ l stock solution in 399 μ l of blocking buffer).
19. Add 150 μ l of goat anti-rabbit secondary antibody onto the fat tissue.
20. Incubate 60 min in a black box with humid paper at RT.
21. Remove the anti-goat secondary antibody solution and wash the cells three times with terrestrial isopod PBS (5 min with 150 μ l of PBS by washing).
22. Mount the preparation in a mixture of DAPI (2.5 mg/ml) to label the nuclei and Citifluor (AF1 antifading).
23. Detection was performed with epifluorescent microscope with apotome equipped with a 63x/1.25 objective (oil immersion) and with the AxioVision 4.8.1 software (Figure 2).

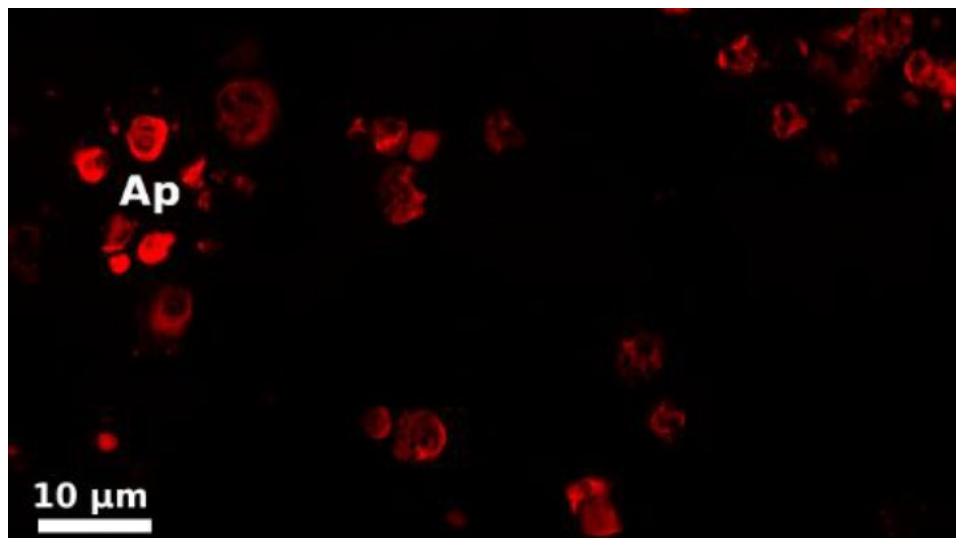


Figure 2. Observation of LC3B labeling in adipocytes collected from *Porcellio dilatatus* infected with the *Wolbachia* strain wVuIC (Le Clec'h et al., 2012).

Recipes

1. Terrestrial isopod PBS
137 mM NaCl
1.5 mM Na₂HPO₄
8 mM KH₂PO₄
3 mM KCl
pH 7.4
2. Blocking buffer
BSA 1% diluted in terrestrial isopod PBS
3. Fixative
3.7% formaldehyde in terrestrial isopod PBS
4. Permeabilization reagen
0.2% Triton X-100 in terrestrial isopod PBS

Acknowledgments

We thank all the technical staff of the UMR EBI CNRS 7267. We also thank Winka Le Clec'h for her valuable comments on the protocol. This work was supported by the Agence Nationale de la Recherche (ADaWOL ANR-09-JCJC-0109-01 coordinated by Mathieu Sicard).

References

1. Chevalier, F., Herbiniere-Gaboreau, J., Bertaux, J., Raimond, M., Morel, F., Bouchon, D., Greve, P. and Braquart-Varnier, C. (2011). [The immune cellular effectors of terrestrial isopod *Armadillidium vulgare*: meeting with their invaders, *Wolbachia*](#). *PLoS One* 6(4): e18531
2. Le Clec'h, W., Braquart-Varnier, C., Raimond, M., Ferdy, J. B., Bouchon, D. and Sicard, M. (2012). [High virulence of *Wolbachia* after host switching: when autophagy hurts](#). *PLoS Pathog* 8(8): e1002844.