

## Primary Culture of Cortical Neuron

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**[Abstract]** Primary culture of neurons from cerebral cortex is a popular model to study neuronal function *in vitro* and to explore the molecular mechanism of neurite outgrowth in the developing and adult central nervous system. This protocol is for preparing a culture of cerebral cortical neurons from postnatal rodent brain (Muramatsu *et al.*, 2012). One day after cell plating, we can observe neurite outgrowth by microscope.

### Materials and Reagents

1. Poly-L-lysine (Sigma-Aldrich, catalog number: P2636)
2. Dulbecco's Modified Eagle Medium, DMEM (Life Technologies, Invitrogen™, catalog number: 12800-017)
3. Fetal bovine serum (Life Technologies, Gibco®, catalog number: 10437)
4. 2.5 % Trypsin solution (Life Technologies, Gibco®, catalog number: 15090-046)
5. DNase (Sigma-Aldrich, catalog number: DN25)
6. Penicillin/streptomycin (Life Technologies, Gibco®, catalog number: 15140)
7. NaH<sub>2</sub>PO<sub>4</sub>·2H<sub>2</sub>O
8. Na<sub>2</sub>HPO<sub>4</sub>·12H<sub>2</sub>O
9. NaCl
10. DMEM
11. FBS
12. Serum-containing culture medium (see Recipes)
13. PBS (see Recipes)

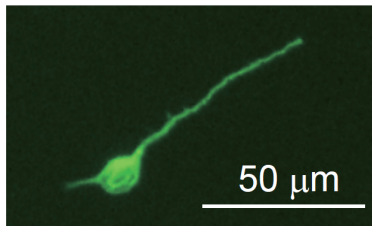
### Equipment

1. Cell culture incubator
2. Centrifuge
3. Stereoscopic microscope

4. Scissors, forceps, and knives
5. Sterile filter (0.45  $\mu\text{m}$ )
6. Cell strainer 70  $\mu\text{m}$  nylon (BD Biosciences, catalog number: 352350)
7. 24 well culture plate

### **Procedure**

1. Coating and washing culture plates.
  - a. Dilute poly-L-lysine stock solution with sterile PBS to 100  $\mu\text{g/ml}$ . Add optimal volume of solution to each well of culture plates. In case of 24 well culture plate, 0.5 ml solution is enough. Coating allows neurons to adhere to the culture plates.
  - b. Leave the plates in incubator for at least 1 h. After that, wash the wells twice with 1 ml/well of sterile PBS. Completely remove the PBS before use. It is not necessary to dry the plate after PBS washing.
2. Choose appropriate anesthesia according to the recommendations of your Institutional Animal Care and Use Committee. After anesthetization, cut the scalp and remove the skull. We use postnatal day 1 mice to measure the length of corticospinal neuron. Pick up whole brain from pups and place it in cold PBS on ice (Chen *et al.*, 2007).
3. Under a microscope, dissect out the cerebral cortex of brains. Remove meninges and blood vessels. The tissues are cut into as small piece (1 mm cubes) as possible with sterile knives.
4. Transfer tissues to a 50 ml tube. The tissues are trypsinized in 10 ml of the trypsin solution (0.25% trypsin, 100  $\mu\text{g/ml}$  DNase in PBS), and are incubated at 37  $^{\circ}\text{C}$  for 15 min.
5. Add an equal amount of 10% FBS-DMEM, and pipettes up and down a few times to help with tissues dissociation.
6. Centrifuge at 300  $\times g$  for 3 min at room temperature, discard the supernatant, and resuspend the pellet in fresh DMEM (not 10% FBS-DMEM).
7. The cells are filtered through a 70  $\mu\text{m}$  nylon cell strainer to obtain single cell suspension. Centrifuge the resulting cell suspension at 300  $\times g$  for 3 min at room temperature. Resuspend the cell pellet in fresh DMEM (not 10% FBS-DMEM).
8. Count the number of cells and adjust the concentration using 10% FBS-DMEM. To measure the neurite length, plate the cells on a 4-well chamber slide at a density of  $5 \times 10^4$  cells/well. Transfer the cell suspension to culture plates and incubate for 24 hours at 37  $^{\circ}\text{C}$  with 5%  $\text{CO}_2$ . To investigate the effect of pharmacological agents, change to the medium containing each agent 2 h after cell plating. Representative image shows the neuronal class III  $\beta$ -Tubulin (Tuj-1) labeled cortical neuron obtained 1 day culture.



**Figure 1. Image of cultured cortical neuron.** Representative image shows the neuronal class III  $\beta$ -Tubulin (Tuj-1) labeled cortical neuron obtained 1 day culture.

### Recipes

1. Poly-L-lysine

Dissolve of sterile distilled water to make stock solution (10 mg/ml). Keep aliquots at  $-20^{\circ}\text{C}$ . Dilute with sterile PBS right before use.

2. Serum-containing culture medium

Reagents	Volume
DMEM	445 ml
FBS	50 ml
Penicillin/streptomycin	5 ml

3. PBS

Reagents	Volume
$\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$	0.312 g
$\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$	2.8652 g
NaCl	8.5 g
DW	1 L

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**References**

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