

In vitro* Floral Induction of *Cuscuta reflexa

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[Abstract] Floral initiation and development in the angiosperms is the essential step on which the yield of the plant depends. Sometimes external climate or any abiotic stress hinders the floral initiation and ultimately affect the plant yield. Hence, *in vitro* floral induction and development can help to overcome the external climatic factor. Furthermore, the protocol for *in vitro* floral induction in the parasitic angiosperm like *Cuscuta reflexa* has not been reported yet. We have standardized the protocol for floral induction in the parasitic plant *Cuscuta reflexa*. In this study it is established that MMS (modified Murashige Skoog) media supplemented with 2 mg L⁻¹ NAA (naphthalene acetic acid, plant growth regulator) and 2 mg L⁻¹ 2,4-D (2,4-dichlorophenoxy acetic acid, plant growth regulator) supported floral induction after shooting *in vitro*. Furthermore, we found that MMS media supplemented with 2 mg L⁻¹ 2,4-D rapidly induced floral buds directly from the nodal explants without any shoot elongation. This protocol will help the researcher to induce flower *in vitro* in the other angiosperm plants along with *Cuscuta reflexa*.

Keywords: *Cuscuta reflexa*, *in vitro*, Floral induction, NAA, 2,4-D

[Background] *Cuscuta reflexa* is a parasitic angiosperm parasitizing on a huge number of angiosperms (Kuijt, 1969). The ability to cause severe damage and loss of yield in the host plant has made this species important for scientific study (Nun and Mayer, 1999). Most of the *Cuscuta* species are non chloroplastic except few which have functional chloroplasts (Hibberd *et al.*, 1998). Development of seedlings from larger embryos of *C. reflexa* in white's medium has been reported (Maheshwari and Baldev, 1961). Floral induction of *C. reflexa* in short day period as well as dark conditions has been seen *in vitro* (Baldev, 1962). *In vitro* floral induction of *Cuscuta japonica* in short day conditions has also been established by Furuhashi *et al.* (1991).

Specifically, floral induction and effect of growth regulators on floral induction of *C. reflexa* has not yet been studied. A complete tissue culture system for *Cuscuta trifolli* in liquid MS culture has been reported so far (Bakos *et al.*, 1995). The floral induction in *C. reflexa* on modified white's medium, subjecting the plant to different light and dark conditions, has been studied years back (Baldev, 1962). Here we have shown a complete *in vitro* culture system for floral induction in *C. reflexa*. Concentration of 2,4-D played a significant role in floral induction. Supplementation of NAA along with 2,4-D induces shoot followed by flower, but supplementation of only 2,4-D induces flower directly from nodal explants without showing any stem elongation. This result is very attractive and is showing the importance of 2,4-D in floral induction of this plant.

Materials and Reagents

1. Pipette tips (Tarsons)
2. 5 ml syringe (Himedia)
3. 0.45 micron filter (Himedia)
4. Stem explants of *Cuscuta reflexa*
5. Murashige & Skoog (MS) basal medium (Murashige and Skoog, 1962) (Sigma-Aldrich, catalog number: M5519)
6. Sucrose (Sigma-Aldrich, catalog number: S0389)
7. Gamborg's vitamin solution (Sigma-Aldrich, catalog number: G1019)
8. Agar (HiMedia Laboratories, catalog number: PCT0901)
9. 2,4-dichlorophenoxyacetic acid (2,4-D) (Sigma-Aldrich, catalog number: D7299)
10. Naphthalene acetic acid (NAA) (Sigma-Aldrich, catalog number: N0640)
11. Sodium hydroxide (NaOH) (Sigma-Aldrich, catalog number: 221465)
12. Mercury(II) chloride (HgCl₂) (0.01%) (EMD Millipore, catalog number: 104419)
13. Bavistin (1% solution, Swat agro chemicals)
14. Ethanol (EMD Millipore, catalog number: 100983)
15. MMS (modified Murashige Skoog) media (see Recipes)
16. 2,4-D stock solution (see Recipes)
17. NAA stock solution (see Recipes)
18. Floral induction media-1 (see Recipes)
19. Floral induction media-2 (see Recipes)
20. HgCl₂ (0.1%) (see Recipes)
21. Bavistin solution (1%) (see Recipes)

Equipment

1. 0.5-10 µl pipette (Transferpette®) (BRAND, catalog number: 704770)
2. 20-200 µl pipette (Transferpette®) (BRAND, catalog number: 704778)
3. 100-1,000 µl pipette (Transferpette®) (BRAND, catalog number: 704780)
4. Beaker (200 ml, Borosil)
5. Laminar hood (Thermo Fisher Scientific)
6. Conical flask (150 ml, Borosil)
7. Culture room (Daihan LabTech India, model: LGC-S201)
8. Autoclave
9. Forceps (ACE Surgical Supply Company, catalog number: 20-000-48)
10. Oven (Hicon India)
11. Water distillation unit (Mars Scientific Instruments, catalog number: BASIC/pH4 & XL)
12. pH meter (CD Hightech, model: APX 175 E)

13. Weighing machine (Sartorius, model: BSA224S-CW)
14. Scissor (general cutting scissor)

Procedure

A. Explant surface sterilization

1. Nodal region (Figure 1A) of the *Cuscuta reflexa* plant (2-month-old) grown naturally on bougainvillea plant was taken as the starting material.
2. Keep small pieces (around 20 pieces taken here) of *Cuscuta reflexa* nodal stem explants (1-2 cm length, Figure 1A) inside a beaker. Treat (submerge) the explants in 100 ml of 1% Bavistin solution for 10 min inside a laminar hood to make those free from dust or any other external contaminations.

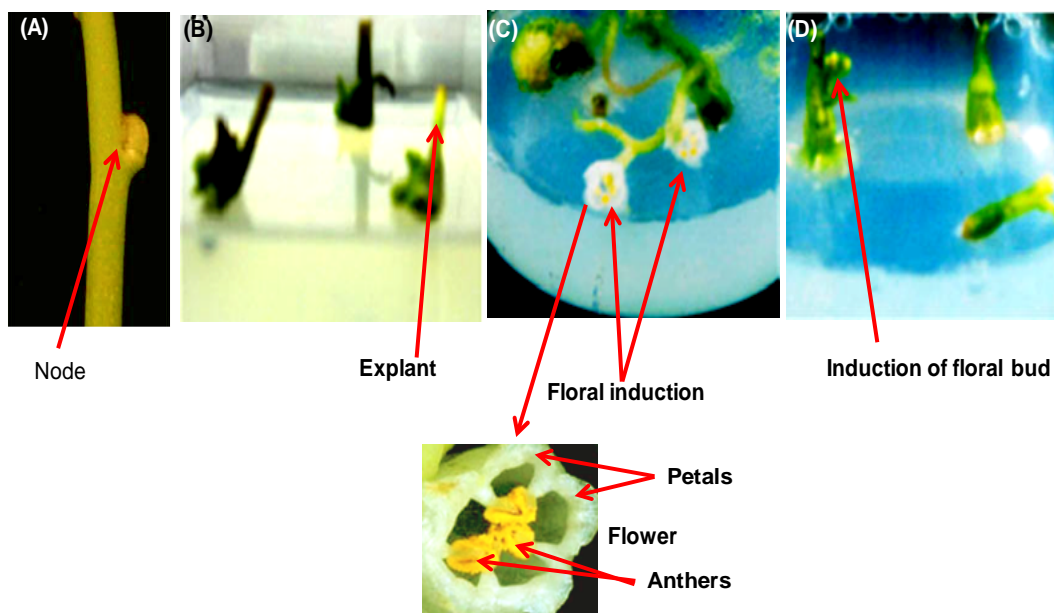


Figure 1. Floral induction from *Cuscuta reflexa* nodal explants. A. Nodal explants inoculated in MMS media supplemented with floral induction hormone/hormones. B and C. Shoot regeneration following floral induction in *Cuscuta reflexa* nodal explants where the MMS media was supplemented with both 2,4-D and NAA. D. Direct floral bud initiation from *Cuscuta reflexa* nodal explants where the MMS media was supplemented with only 2,4-D.

3. Rinse the explants with autoclaved distilled water inside a laminar hood by brief shaking with hand.
4. Decant the distilled water and pour 100 ml of 0.1% HgCl₂ into the same beaker for surface sterilization inside the laminar hood. Surface sterile the explants for 5 min with brief shaking by hand.

B. Inoculation of explants for floral induction

1. Inoculate some of the explants (stuck up to 0.5 cm of 3 to 4 explants into media of one flask, Figure 1A) in the conical flask containing floral induction media-1 and inoculate some explants in floral induction media-2 (method for preparation of floral induction media-1 and -2 is described in Recipes section).
2. Plug all the conical flasks containing the media with the explants air tight to avoid any bacterial and/or fungal contamination.
3. Keep the conical flasks inside a tissue culture room where controlled light (1,000 lux) and temperature condition (28 ± 1 °C, 12 h dark/light period) will be maintained.
4. Notice the proliferation of nodal region (Figure 1B) and generation of shoots and/or floral buds. It will take around 25 to 30 days to initiate floral buds.
5. Floral induction media-1 (200 μ l 2,4-D/L and 200 μ l NAA/L supplemented in MS media) results in shoot regeneration following flowering initiation (Figure 1C). But floral induction media-2 (200 μ l 2,4-D/L supplemented in MS media) resulted in floral bud initiation without shoot elongation (Figure 1D).

Data analysis

The experiment has been repeated thrice independently. All the analysis has been done manually or visually.

Notes

1. Nodal region of the plant should be taken as explants as from the nodal region new callus or branch will develop quickly in comparison with the other parts.
2. Hands should be washed and sterilized (with 70% ethanol) properly during the whole experimental process.
3. Hormones should be filter sterilized before use.
4. Floral induction media should be prepared in the conical flasks or jam bottles instead of Petri plates to get some space for the development of vine shoots.
5. Surface sterilization of the explants should not exceed more than 10 min as it may cause tissue damage.
6. Light, temperature and humidity should be maintained properly in the culture room.
7. If bacterial or fungal contamination is observed on the media surface, the explants should be transferred to new flasks/jam bottles containing fresh media.
8. 2,4-D and NAA can also be added before autoclaving of media. But if filter sterilized, it can be added after autoclaving of media. The media need not to be pre-cooled for adding hormones.

Recipes

1. MMS media (modified MS media containing basal MS media with vitamins of Gamborg's media [Gamborg *et al.*, 1968])
 - a. Weigh 4.4 mg of readily available MS powder
 - b. Dissolve 4.4 mg of MS powder in 1 L of distilled water. And then add 30 g of sucrose to 1 L of MS solution
 - c. Add 1 ml of Gamborg's vitamin solution to 1 L of MS basal solution
 - d. Adjust pH to 5.8 with 1 N KOH
 - e. Add 8 g of agar to the media and autoclave for 15 min at 121 °C
2. 2,4-D stock solution
 Dissolve 10 mg of 2,4-D in 200 µl of ethanol
 Add 800 µl of distilled water to get 1 ml final volume
 Then filter sterilize.
3. NAA stock solution
 Dissolve 10 mg of NAA in 200 µl of 1 N NaOH
 Add 800 µl of distilled water to get 1 ml final volume
 Then filter sterilize
4. Floral induction media-1
 Add 2 mg of filter sterilized 2,4-D and 2 mg of NAA to 1 L of MMS media after autoclaving
5. Floral induction media-2
 Add 2 mg of filter sterilized 2,4-D to 1 L of MMS media after autoclaving
6. HgCl₂ (0.1%)
 Dissolve 0.1 mg HgCl₂ in 100 ml of autoclaved distilled water
7. Bavistin solution (1%)
 Dissolve 1 g of Bavistin in 100 ml of autoclaved distilled water

Note: All the solutions can be freshly prepared or stored solutions (at 4 °C, stable for 30 days).

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