

## A Simple RNA Preparation Procedure from Yeast for Northern Blot Using Hot Phenol

Yuehua Wei\*

Department of Pharmacology, Cancer Institute of New Jersey, UMDNJ Robert Wood Johnson Medical School, Piscataway, USA

\*For correspondence: [weiyh.sju.edu@gmail.com](mailto:weiyh.sju.edu@gmail.com)

**[Abstract]** Compared to several expensive RNA extraction kits, the following protocol provides an economic and simple method for researchers to extract yeast RNA. This method can achieve RNA quality that is sufficient for most northern blot studies in yeast.

### Materials and Reagents

1. W303a cell line
2. Sodium acetate trihydrate ( $\text{CH}_3\text{CO}_2\text{Na}\cdot 3\text{H}_2\text{O}$ ) (Sigma-Aldrich, catalog number: 236500)
3. Phenol ( $\text{C}_6\text{H}_5\text{OH}$ ) (Sigma-Aldrich, catalog number: P1037)
4. EDTA ( $\text{Na}_2\text{EDTA}\cdot 2\text{H}_2\text{O}$ ) (Sigma-Aldrich, catalog number: ED2SS)
5. Acetic acid ( $\text{CH}_3\text{COOH}$ ) (Sigma-Aldrich, catalog number: 320099)
6. Chloroform ( $\text{CHCl}_3$ ) (Sigma-Aldrich, catalog number: 472476)
7. 8-hydroxyquinoline ( $\text{C}_9\text{H}_7\text{NO}$ ) (Sigma-Aldrich, catalog number: 252565)
8. NaCl (Thermo Fisher Scientific, catalog number: S641-500)
9. Synthetic complete (SC) medium
10. SDS (Sigma-Aldrich, catalog number: L3771)
11. Isoamyl alcohol (Sigma-Aldrich, catalog number: W205710)
12. Ethanol (Thermo Fisher Scientific, catalog number: 64-17-5)
13. DEPC water
14. Phenol-AE buffer (see Recipes)
15. Phenol:  $\text{CHCl}_3$  /AE-Na (see Recipes)
16.  $\text{CHCl}_3$ : Isoamyl alcohol (24:1) (see Recipes)

### Equipment

1. Standard bench-top centrifuge
2. Microfuge
3. Shaker
4. 1.5 eppendorf tube

## 5. Liquid nitrogen

**Procedure**

1. Prepare reagents according to recipes. Note that everything is in DEPC water.
2. Inoculate W303a cells expressing different TOR1-RR variants in 2 ml SC medium overnight.
3. Subculture the cells starting from  $OD_{600}=0.1$  in 10 ml SC media, shake vigorously at 30 °C, 300 rpm for around 4-6 h until  $OD_{600}=0.4-0.5$ .
4. Collect the cells by spinning down without freezing on ice. Discard supernatant.
5. Re-suspend cells with 1 ml water and transfer to a 1.5 eppendorf tube, quickly spin down at 3,000 x *g* for 15 sec.
6. Re-suspend cell pellet in 400  $\mu$ l of AE buffer at room temperature.
7. Add 40  $\mu$ L 10% SDS (final around 1%) and vortex briefly at room temperature (RT).
8. Immediately add 500  $\mu$ l hot phenol/AE (put in 65 °C for 10 min before use), vortex vigorously for 1 min.
9. Incubate at 65 °C for 5 min. Briefly vortex every 30 sec.
10. Immediately freeze by dumping into liquid nitrogen (labels won't get off).
11. Wait to thaw at RT (put in 30 °C to thaw may crack the tube).
12. Centrifuge for 10 min on a standard laboratory microfuge at 20,000 x *g* at RT.
13. Transfer around 400  $\mu$ l supernatant to a new eppendorf tube. Recycle the lower phenol fraction carefully following the chemical safety protocol in your laboratory.
14. Add equal volume (400  $\mu$ l) phenol:  $CHCl_3$ /AE-Na. Vortex vigorously for 1 min at RT.
15. Spin down at 20,000 x *g* for 5 min in a standard laboratory microfuge.
16. Transfer supernatant (around 350  $\mu$ l) to a fresh 1.5 ml eppendorf tube.
17. Add  $CHCl_3$ : isoamyl alcohol (24:1). Vortex vigorously for 1 min at RT.
18. Transfer aqueous supernatant to fresh 1.5 ml microfuge tube. If white cloudy precipitate is observed between aqueous phase and organic phase, repeat steps 17-18.
19. Add 1/10 volume of 3 M NaOAc (pH 5) and vortex vigorously. Add 2.5 volumes of ethanol. Vortex again.
20. Place at -20 °C for at least 30 min.
21. Spin down in the microfuge at 20,000 x *g*, 15 min at 4 °C. RNA pellet is usually visible.
22. Add ice-cold 75% EtOH, place at 4 °C for around 10 min. Vortex and spin down on microfuge 20,000 x *g*, 15 min at 4 °C.
23. Discard supernatant. Suck out the liquid droplets in the tube.
24. The white RNA pellet will turn clear when it dries out. Add 30-50  $\mu$ l ddH<sub>2</sub>O (DEPC) immediately after it becomes clear.

25. Do not let the RNA over-dry, which will make it difficult to dissolve. If RNA pellet is over-dry, dissolve RNA at 37 °C for 30 min.
26. Store RNAs at -80 °C for more than 2 months.

### **Recipes**

1. Phenol-AE buffer  
Make Acetate-EDTA(AE) buffer  
50 mM NaOAc  
10 mM EDTA (pH 5.0)  
Liquefied phenol equilibrated with equal volume of AE buffer. Store at 4 °C.
2. Phenol: CHCl<sub>3</sub> /AE-Na  
Make AE-Na buffer  
10 mM NaOAc  
2 mM EDTA  
100 mM NaCl (pH 6.0)  
50% phenol/AE  
50% CHCl<sub>3</sub>  
0.25% 8-hydroxyquinoline  
Store at 4 °C.
3. CHCl<sub>3</sub>: Isoamyl alcohol (24:1)

### **Acknowledgments**

This protocol was adapted from and used in Wei and Zheng (2009) and Wei *et al.* (2009).

### **References**

1. Wei, Y. and Zheng, X. F. (2009). [Sch9 partially mediates TORC1 signaling to control ribosomal RNA synthesis](#). *Cell Cycle* 8(24): 4085-4090.
2. Wei, Y., Tsang, C. K. and Zheng, X. F. (2009). [Mechanisms of regulation of RNA polymerase III-dependent transcription by TORC1](#). *EMBO J* 28(15): 2220-2230.