

Dye Labeling of Live *Nippostrongylus brasiliensis* Larvae for Visualization in Host Tissue

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[Abstract] Visualization of the interaction between parasitic nematodes and their host enables a better understanding of the development of the nematode during the infectious stages of its life cycle and of the effects of host response on nematode integrity in tissues. Appropriate live imaging of these nematode/host interactions, to date has been hindered by the lack of appropriate molecular tools or efficient labeling agents. Here, we present techniques for the live labeling of the nematode parasite *Nippostrongylus brasiliensis* (*N. brasiliensis*) that allows visualization of the parasite in the mouse host for up to 24 h. The external sheath can be labeled with CFSE allowing infective larvae to be identified and followed until the stage of exsheathment. The internal labeling of infective parasites can be performed by ingestion of NY microspheres. The worms can continue to be identified for up to 24 h following exsheathment. This should be applicable to other parasitic nematodes.

Materials and Reagents

1. 200 µl pipette tips (Corning, Axygen[®], catalog number: T-300-L-R-S)
2. 10 µl pipette tips (Corning, Axygen[®], catalog number: T-300-R-S)
3. 1 ml pipette tips (Corning, Axygen[®], catalog number: T-1000-C-L-R-S)
4. 1.7 ml MaxyClear Snaplock Microcentrifuge Tube (Corning, Axygen[®], catalog number: MCT-175-C)
5. 24-well Tissue culture plate (Thermo Fisher Scientific, Falcon[™], catalog number: 353047/08-772-1)
6. Insulin syringe 29 G, 0.3 ml ultrafine (BD, catalog number: 320431)
7. 15 ml Conical Centrifuge Tubes (Thermo Fisher Scientific, Falcon[™], catalog number: 352096/14-959-49B)
8. Microscope slides (Thermo Fisher Scientific, catalog number: LBS2951RC)
9. Petri dish, 60 mm x 15 mm (In Vitro Technologies, catalog number: FAL351007)
10. Tin-foil (any grocery store)
11. *N. brasiliensis* L3 infective larvae (iL3) (Camberis *et al.*, 2003)

Note: Camberis et al. (2003) provides a detailed review on the preparation of N. brasiliensis.

Note: There are several labs that maintain the life cycle and can be contacted through their publication address.

12. C57Bl/6J Mouse (or strain of choice)
13. Fluoresbrite® YO Carboxylate Microspheres 0.50 µm (Polysciences, catalog number: 18720)
14. 5(6)-CFDA, SE; CFSE (5-(and-6)-Carboxyfluorescein Diacetate, Succinimidyl Ester), mixed isomers (Life Technologies, Molecular Probes™, catalog number: C1157)
Note: Currently, it is "Thermo Fisher Scientific, Molecular Probes™", catalog number: C1157".
15. DMEM, high glucose, pyruvate (Life Technologies, Gibco™, catalog number: 11995-065)
Note: Currently, it is "Thermo Fisher Scientific, Gibco™", catalog number: 11995-065".
16. PBS (DPBS, no calcium, no magnesium) (Life Technologies, Gibco™, catalog number: 14190-144)
Note: Currently, it is "Thermo Fisher Scientific, Gibco™", catalog number: 14190-144".
17. Penicillin/Streptomycin 10,000 U/ml (Life Technologies, Gibco™, catalog number: 15140-122)
Note: Currently, it is "Thermo Fisher Scientific, Gibco™", catalog number: 15140-122".
18. Gentamicin solution 10 mg/ml (Sigma-Aldrich, catalog number: G1272)
19. Fetal Calf Serum, qualified, US origin (Life Technologies, Gibco™, catalog number: 26140-079)
Note: Currently, it is "Thermo Fisher Scientific, Gibco™", catalog number: 26140-079".
20. TWEEN® 20 (Sigma-Aldrich, catalog number: P2287)
21. Dimethyl sulfoxide (DMSO) (Sigma-Aldrich, catalog number: D2650)
22. Wash solution I (see Recipes)
23. Wash solution II (see Recipes)
24. Culture Solution (see Recipes)
25. CFSE (see Recipes)

Equipment

1. Stereo-microscope with fluorescence capabilities (Olympus, model: SZX16)
2. Confocal microscope for detailed analysis (Olympus FV1200 Scanning Laser Confocal Microscope) (if required)
3. HeraCELL 150 CO₂ Incubator set at 37 °C, 5% CO₂ (Thermo Fisher Scientific, model: NC0689918)
4. Pipettes (P10, P200 and P1000) (Gilson)
5. Iris scissors (Medicon, catalog number: 202.20.11)
6. Forceps (Medicon, catalog number: 07.55.20)

Procedure

If only external labeling of the larvae is required, follow step A1 from the Internal labeling protocol and then proceed with the external labeling protocol.

A. Internal labeling of *N. brasiliensis* iL3 using YO Microspheres

1. Prepare sufficient *Nippostrongylus brasiliensis* L3 larvae for experiment by washing larvae three times with 3 ml of PBS in a 15 ml tube and allowing larvae to settle by gravity after each wash. Allowing the larvae to settle by gravity should take approximately 15 min and the larvae should form a pellet at the bottom of the 15 ml tube. This is carried out at room temperature.

Note: 600 Nippostrongylus brasiliensis L3 larvae are used per injection. By preparing at least 700 Nippostrongylus L3 per injection, it allows for any loss during the labeling procedure.

2. Transfer the pellet of L3 into a 1.7 ml micro-centrifuge tube using a 1 ml pipette, allow the larvae to form a pellet at the bottom of the tube by gravity for approximately 15 min and then discard any remaining PBS.
3. Add 500 μ l of Wash solution I and incubate at 37 °C in a water-bath for 1 h. Following incubation, discard Wash solution I using a pipette.
4. Prepare a suspension of 1,500 L3 per ml in Culture Solution by microscopically counting a 10 μ l sample of the L3 suspension and adjusting the concentration of larvae with Culture Solution.
5. Add 1 ml of 1,500 L3 per ml suspension into wells of a 24 well plate. Incubate at 37 °C, 5 % CO₂ overnight.
6. Add 1 μ l of YO microspheres to each well and mix by gently swirling the plate.
Please refer to Note 1 in the Notes section for more information regarding this step.
7. Incubate for 4 h at room temperature in the dark by placing in a drawer or covering the plate with tinfoil.
Please refer to Notes 2-3 in the Notes section for more information regarding this step.
8. Add 1 ml Wash solution II to each well and mix by gently swirling the plate.
9. Transfer the larvae from each well into 15 ml tubes covered with tin foil to protect from light. Let the larvae settle for approximately 15 min to form a pellet and discard supernatant.
10. Add 3 ml Wash Solution II to each tube and allow larvae to settle by gravity.
11. Discard supernatant and repeat at least three times or until the supernatant becomes clear.

B. External labeling of *N. brasiliensis* iL3 using CFSE

1. Transfer the larval pellet in a 1.7 ml micro-centrifuge tube and cover with tin foil to protect from light.

2. Allow the larvae to settle and discard supernatant using a pipette.
3. Add 1 μ l of 10 mM CFSE to the pellet and mix by gently flicking the micro-centrifuge tube 2-3 times.
4. Incubate at room temperature for 8 min.
5. Add 1 ml Wash solution II and mix by inverting 2-3 times.
6. Allow larvae to settle by gravity and discard supernatant using a pipette.
7. Repeat wash step at least three times or until the supernatant becomes clear.

C. *In vivo* imaging of *N. brasiliensis* iL3

1. Prepare a suspension of 600 L3 per 30 μ l in PBS by microscopically counting a 10 μ l sample of the L3 suspension and adjusting the larval concentration with PBS.
2. Using an insulin syringe and needle, inject 30 μ l of the L3 suspension in the appropriate site of the body intended for study:
 - a. Use intra-dermal (i.d.) administration of *N. brasiliensis* iL3 on the ventral surface (belly skin) or the ear pinnae for imaging of the skin. Hair should be plucked around the site of injection for optimal imaging results (Figure 1).
 - b. Use intra-venous (i.v.) administration of *N. brasiliensis* iL3 for imaging of worms migrating to lung.

Note: Intra-nasal administration of live larvae does not lead to direct infection and development in the lung. Rather, the infective larvae must first pass into the venous blood supply from the airway mucosa and arrive in the lungs 16-18 h later.



Figure 1. Representative image of intra-dermal administration of *N. brasiliensis* larvae in the ear pinna with India ink incorporated in the injected solution for visualization purposes only [adapted from Camberis *et al.* (2013)]

3. Harvest the tissue of interest
 - a. Ear-Euthanise the mouse using an approved form of euthanasia, remove the pinna using scissors and place in a Petri dish containing cold PBS for approximately 15 min to lightly anaesthetize the larvae.
 - b. Belly skin-Euthanise the mouse using an approved form of euthanasia, cut a 1 cm x 1 cm piece of skin around the injection site using scissors, peel skin off and

place the skin piece ventral side down in a Petri dish containing cold PBS for approximately 15 min to lightly anaesthetize the larvae. Take care to spread the skin out.

- c. Lung-Euthanise the mouse using an approved form of euthanasia. Expose the thoracic cavity and remove the lungs in one piece and place in a Petri dish containing cold PBS for approximately 15 min to lightly anaesthetize the larvae.
4. Transfer the tissue onto a glass slide.
 - a. Belly skin-Place a cover slip onto the tissue.
 - b. Ear-Flatten the ear pinna as much as possible.
 - c. Lung-Place lung ventral side up.
5. Image using fluorescent stereomicroscope or Confocal microscope for more detailed imaging in the tissue.

For visualizing dyes using the Confocal microscope:

CFSE 473 nm laser -BP 515/50

NY 559 nm laser-BP 625/100

For visualizing dyes using the stereo microscope with fluorescence:

CFSE BP Excitation filter 495/10-Emission BP 538/45

NY Excitation filter 545/30-Emission filter BP 610/75

Representative data

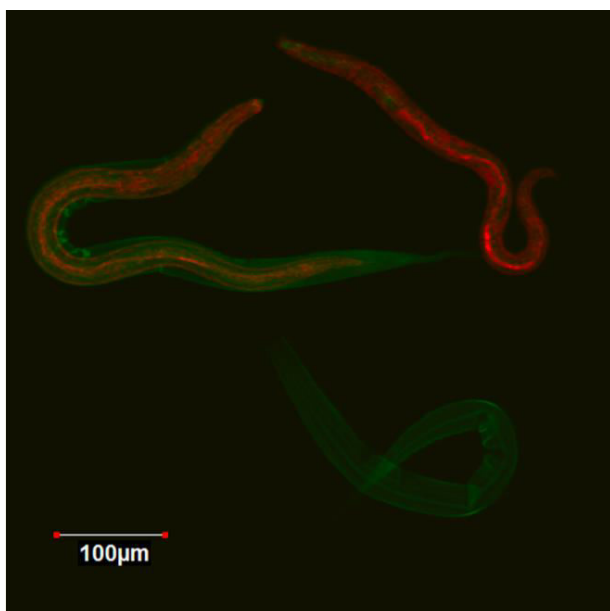


Figure 2. Representative Fluorescent microscopy image showing: Top left, *Nippostrongylus* larvae stained internally with YO microspheres and stained externally with CFSE, Top right, *Nippostrongylus* larvae stained internally with YO micro beads and Bottom right, CFSE labeled empty larval sheath

Notes

Two thorough reviews of the basic techniques involved in the preparation, handling and use of *N. brasiliensis* are Kassai (1982) and Camberis *et al.* (2003).

The protocol outlined above enables reproducible staining of nematodes and their visualization *in vivo* in tissues for up to 24 h.

1. *N. brasiliensis* larvae need to be feeding for the ingestion of fluorescent YO microspheres. In order to activate feeding, larvae need to be incubated at 37 °C in 5% CO₂ for a 10-h incubation period. Incubation periods of six hours or less are insufficient to activate the larvae to feed.
2. The microspheres used in this protocol are fluorescent polystyrene microspheres that have carboxylate groups on their surfaces, which can aid in the covalent coupling of proteins. They can be toxic to the larvae; both the volume of microspheres and incubation times are critical factors in determining whether the larvae will survive. Microsphere volumes greater than 1 µl microspheres are toxic and should be avoided.
3. The four-hour incubation period of the microspheres with the larvae has been optimized to allow for a signal bright enough for *in vivo* visualization with minimal toxicity, as larvae can still complete the life cycle through to fecund adults in the gut. In our hands, incubation times greater than 4 h are toxic, however, incubation times less than 4 h do not provide a bright enough signal for *in vivo* visualization.

Recipes

1. Wash solution I
 - 100 ml PBS
 - 5 ml Penicillin Streptomycin
 - 1.5 ml Gentamicin solution
 - Can be made in advance and stored for up to one month at 4 °C
2. Wash solution II
 - 100 ml PBS
 - 0.05 ml Tween 20
 - Can be made in advance and stored for up to one month at 4 °C
3. Culture solution
 - 100 ml DMEM
 - 10 ml fetal calf serum
 - 5 ml Penicillin Streptomycin
 - Can be made in advance and stored for up to one month at 4 °C
4. CFSE
 - 10 mM CFSE in DMSO
 - 5.57 mg CFSE in DMSO (1 ml)

Stored frozen in 10 µl aliquots

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