

## **Chitin-challenged Mice Model to Study M2 Macrophages Polarization**

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**[Abstract]** Chitin is a key component of insects, fungi, and house-dust mites. Chitin has been shown to induce M2-type immune responses *in vivo*. Intranasal or intraperitoneal (i.p.) administration of chitin particles results in infiltration of eosinophils to the local sites and activation of macrophages with a M2 phenotype. Chitin-challenged mice model can be used to induce M2 macrophages polarization and thus to analyze the M2 phenotype from isolated peritoneal cells.

### **Materials and Reagents**

1. Mice C57BL/6 male (age 8-12 weeks)
2. Chitin (Sigma-Aldrich, catalog number: C9752)
3. 70% ethanol
4. PBS (Lonza, catalog number: BE 17-515Q)
5. 70  $\mu$ m sterile cell strainers (BD Biosciences, catalog number: 352350)
6. RPMI 1640 (Lonza, catalog number: BE 12-115F)
7. 10 ml syringe
8. 21 G Needles
9. 25 G Needles
10. 15 ml conical tubes

### **Equipment**

1. Sonication bath, P-selecta ultrasons (150 W)
2. Scissors
3. Bench-top refrigerated centrifuge

### **Procedure**

#### **A. Chitin administration**

1. Preparation of chitin
  - a. Weight 800 ng of chitin per mouse and wash 3 times with PBS (5 min, 13,000 rpm, 16,800  $\times$  g).

- b. Suspend chitin in 1 ml of PBS and sonicate in bath for 1 h [at the maximum setting of the *sonicator* (Ultrasons, Selecta)].
 

*Note: Alternatively chitin can be sonicated at 25% output power three times for 5 min with a Branson sonicator.*
  - c. Filter the suspension with 70 µm sterile cell strainers.
2. Inject 1 ml of chitin suspension i.p. per mouse using a 10 ml syringe with a 25 G needle. Wait for 2 days and harvest peritoneal cells.
- B. Isolation of chitin elicited peritoneal exudate cells (PECs)
1. Sacrifice mouse with CO<sub>2</sub> or isofluorane (the use of cervical dislocation is not indicated in this protocol in order to avoid potential internal bleeding).
  2. Clean abdomen with 70% ethanol.
  3. Remove skin to expose the peritoneal wall. Practice a small incision in the skin and pull firmly.
  4. Inject 10 ml of RPMI 1640 into the peritoneal cavity with a 25 G needle.
  5. Massage abdomen for approximately 30 sec.
  6. Recover peritoneal fluid as much as possible using a 10 ml syringe with a 21G needle. (Usually approximately 8-10 ml fluid can be recovered from one mouse.)
  7. Remove needle from syringe and put fluid into a 15 ml conical centrifuge tube on ice.
  8. Centrifuge peritoneal cells 5 min at 300 x g (1,500 rpm, 4 °C). Discard the supernatant and collect the pellet.
  9. Resuspend cell pellet in 10 ml of RPMI 1640 medium and count cells. (The cell number expected to be recovered is about 5-8 millions.)
  10. Analyze peritoneal cells to confirm M2 activation by performing either real-time PCR or flow cytometry techniques to detect typical M2 markers (*e.g.* Arginase-1, Ym-1, Fizz-1, mannose receptor).

### **Acknowledgments**

This study was supported by grant PI11.0036 from the FIS and MPY 1410/09 from ISCIII to SH, and by grant TPY-M-1068/13 to AL. AL was supported by MINECO-ISCIII, through the Miguel Servet Programme (CP12/03087). L. J-G. was supported by FIS (FI12/00340). This protocol is adapted from Jimenez-Garcia *et al.* (2015).

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