

## Substrate Specificity of Recombinant Ser/Thr Protein Kinase

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**[Abstract]** Protein kinases are enzymes that phosphorylate proteins in a cell. Determination of kinase activity in reactions of phosphorylation is a very convenient way for a biochemical characterization of this group of enzymes. Here we describe a method to determine the activity of a recombinant Ser/Thr protein kinase using as a possible substrate MBP, H1, and BSA.

### **Materials and Reagents**

1. *Escherichia coli* (*E.coli*) strains - for cloning and for expression - DH5 $\alpha$ , Rosetta (DE3 pLysS)
2. Specific synthetic oligonucleotides
3. PCR reagents (dNTP, MgCl<sub>2</sub>, PCR buffer, Taq-polymerase, DNA) (Thermo Fisher Scientific, catalog numbers: R0181, EP0612)
4. Plasmids for cloning and for expression [pTZ57R/T, pET41a(+)]
5. CaCl<sub>2</sub> (Sigma-Aldrich, catalog number: C3306-250G) (for transformation)
6. Antibiotics (Ampicillin, Kanamycine, Chloramphenicol) (AppliChem, catalog numbers: A-0839, A-1493, A-6435)
7. Isopropyl thio- $\beta$ -D-galactoside (IPTG) (Fermentas, catalog number: R0393)
8. Tris-HCl (Sigma-Aldrich, catalog number: T6791)
9. Ethylenediaminetetraacetic acid (EDTA) (AppliChem, catalog number: A-1104.0500)
10. Dithiothreitol (DTT) (Sigma-Aldrich, catalog number: 43815-5G)
11. Ethylene glycol-bis(2-aminoethylether)-N,N,N',N'-tetraacetic acid (EGTA) (Sigma-Aldrich, catalog number: E3889-10G)
12. Na<sub>3</sub>VO<sub>4</sub> (Sigma-Aldrich, catalog number: S6508-10G)
13. NaF (Sigma-Aldrich, catalog number: S7920-100G)
14.  $\beta$ -glycerophosphate (Sigma-Aldrich, catalog number: G9422-50G)
15. Sucrose (AppliChem, catalog number: A-1125)
16. Phenylmethylsulfonyl fluoride (PMSF) (AppliChem, catalog number: A-0999)
17. Benzamidine hydrochloride (Sigma-Aldrich, catalog number: B6506)
18. 6-aminocaproic acid (Sigma-Aldrich, catalog number: A7824-25G)
19. 2-(N-morpholino)ethanesulfonic acid (MES) (Amresco, catalog number: Am-E169-0.05)
20. Myelin Basic Protein bovine (MBP) (Sigma-Aldrich, catalog number: M1891-5MG)
21. Histone from calf thymus (H1) (Sigma-Aldrich, catalog number: H5505-100MG)

- 38 22. Bovine serum albumin (BSA) (Sigma-Aldrich, catalog number: 05470-1G)
- 39 23. MgCl<sub>2</sub> (AppliChem, catalog number: A-1447)
- 40 24. Brilliant Blue R Concentrate (Sigma-Aldrich, catalog number: B8647)
- 41 25. BCA Protein Assay Kit (Pierce Thermo Scientific, catalog number: 23227)
- 42 26. Sodium dodecyl sulfate, SDS (Sigma-Aldrich, catalog number: L3771)
- 43 27. TGX FastCast Acrylamide Starter Kit (12%) (Bio-Rad Laboratories, catalog number: 161-0175)
- 44 28. N,N,N',N'-tetramethylethylenediamine (TEMED) (Bio-Rad Laboratories, catalog number: 161-
- 45 0800)
- 46 29. Ammonium Persulfate (APS) (Bio-Rad Laboratories, catalog number: 161-0700)
- 47 30. Illustra NAP-5 Columns (GE Healthcare Life Sciences, catalog number: 17-0853-02)
- 48 31. [ $\gamma$ -<sup>32</sup>P] ATP
- 49 32. Homogenization buffer (see Recipes)
- 50 33. Reaction mixture (see Recipes)

51

## 52 **Equipment**

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- 54 1. Temperature controlled shaker (Heidolph, model: Unimax 1010)
- 55 2. French Pressure Cell Press (Aminco) or glass beads (Sigma-Aldrich, catalog number: G4649) for
- 56 cell disruption
- 57 3. Thermostat
- 58 4. Spectrophotometer
- 59 5. Gel electrophoresis equipment (Mini-PROTEAN Tetra Handcast Systems, Bio-Rad Laboratories,
- 60 catalog number: 165-8000)
- 61 6. Laminar flow box
- 62 7. Refrigerated microcentrifuge preset to 4 °C (Eppendorf, model: 5415 R)
- 63 8. Gel Dryer System (Bio-Rad, catalog number: 165-1789)
- 64 9. Exposure cassette/BioMax<sup>®</sup> intensifying screen set (Sigma-Aldrich, catalog numbers: C4979 and
- 65 P8432)
- 66 10. Carestream<sup>®</sup> Kodak<sup>®</sup> autoradiography GBX developer/replenisher (Sigma-Aldrich, catalog
- 67 number: P7042)
- 68 11. Carestream<sup>®</sup> Kodak<sup>®</sup> autoradiography GBX fixer/replenisher (Sigma-Aldrich, catalog number:
- 69 P7167)

70

## 71 **Procedure**

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- 73 A. Expression and isolation of a recombinant protein kinase

- 74 1. Amplify a DNA fragment carrying the gene of interest using synthetic primers. For the more  
75 convenient cloning of the desired insert, it is worth to construct the primers harboring specific  
76 restriction sites for endonucleases. The easiest way to clone this PCR product is to use T-vector  
77 pTZ57R/T. Transform DH5- $\alpha$  strain. Spread transformed cell onto LB agar plates containing  
78 ampicillin, IPTG, X-GAL. Incubate overnight at 37 °C. Select white colonies that should harbor  
79 plasmid with a desired insert and inoculate 3 ml of LB-medium. Extract pDNA from the overnight  
80 culture. Check it for containing your full length insert either via PCR or via restriction analysis.
- 81 2. Ligate your restriction fragment with the appropriate expression vector.
- 82 3. Transform *E. coli* expression strain with the resultant plasmid using CaCl<sub>2</sub> method (Sambrook *et*  
83 *al.*, 1989). Spread transformed cells on the LB-plate containing specific antibiotic that is coded by  
84 the expression plasmid. Manipulations with bacterial cells should be carried out in a flow box (He,  
85 2011).
- 86 4. Grow the transformed cells in 100 ml of LB-medium until the OD<sub>595</sub> of suspension reached 0.5.  
87 Initiate overproduction of the recombinant protein by the addition of IPTG to a final concentration  
88 of 0.1 mM. Induction conditions may vary strongly (from 3 h at 37 °C to 15 h at 25 °C). Centrifuge  
89 cells at 3,500 x *g* for 10 min at 4 °C.
- 90 5. Resuspend the collected cells in a prechilled homogenization buffer (1 g of cells in up to 5 ml of  
91 buffer). Disrupt the cells either using a French Pressure Cell Press (2.8 MPa) or glass beads (use  
92 an equal volume of beads and cell suspension. Vortex vigorously for 30 sec. To avoid warming of  
93 the sample incubate a test tube for 30 sec on ice. Repeat this procedure 10 times). All  
94 procedures should be carried out at 4 °C.
- 95 6. Centrifuge the crude extracts stepwise, first at 7,500 x *g* for 15 min to remove unbroken cells and  
96 afterwards at 16,000 x *g* for 20 min for clarifying cell lysate. Procedures should be carried out at  
97 4 °C. Collect supernatant (soluble protein fraction) and keep a pellet.
- 98 7. Desalt the soluble protein fraction on NAP-5 columns according manufacturer's instruction. Elute  
99 the protein with 20 mM MES-KOH (pH 6.5) and determine protein concentration using BCA  
100 Protein Assay Kit. To determination protein quantity in the pellet resuspend it with 4% of SDS and  
101 recover the proteins by centrifugation at 16,000 x *g* for 20 min. Then determine protein  
102 concentration with BCA Protein Assay Kit. Mix protein sample with appropriate amount of  
103 Laemmli sample buffer to get the final protein concentration 1 mg/ml (He, 2011).
- 104 8. Prepare and run SDS-PAGE. Stain the gel with CBB R250 (Yan, 2011).
- 105 9. Analyze both the desalted soluble fraction and proteins from the pellet using SDS-PAGE. If your  
106 protein is soluble proceed to the protein kinase assay. If the protein is in the pellet, it is necessary  
107 to recover the protein from inclusion bodies or to start over induction procedure in other  
108 conditions, trying to shift the protein into the supernatant. Alternatively, purify the soluble proteins  
109 further by affinity chromatography. The first possibility allows to obtain the result much quickly.

110 The second one requires more time and an additional purification step but gives the better picture  
111 with the fewer non specific bands.

112

### 113 B. Phosphorylation *in vitro*

114 1. Kinase activity assay is performed in a reaction mixture containing 20 mM MES-KOH (pH 6.5), 10  
115 mM MgCl<sub>2</sub>, 10 μM ATP, exogenous protein substrate, 74 kBq 2μCi [γ-<sup>32</sup>P] ATP (specific activity  
116 148 TBq/mmol). The quantity of the labeled ATP depends on the quantity of the protein. Different  
117 proteins (histone H1, basic myelin protein MBP, and BSA) can be used as possible exogenous  
118 substrates. To calculate the volume of the [γ-<sup>32</sup>P] ATP required you can use, for example, this site  
119 <http://www.radprocalculator.com/Decay.aspx>. Or you can do it manually:

120 Radioactive concentration (μCi/μl) = Calibration Concentration (μCi)/2<sup>N/14.3</sup>,

121 where N - number of days passed from the day of calibration, taken from the passport of the  
122 product. 14.3 days - half-life period of the <sup>32</sup>P.

123 The reaction is initiated by the addition of 5-100 μg of the recombinant protein kinase and allowed  
124 to proceed for 15 min at 30 °C. The quantity of the recombinant protein kinase in this reaction  
125 depends on whether affinity purified or non-purified the recombinant protein kinase is used.

126 2. The reaction is terminated by adding half of the reaction volume of 3x Laemmli sample buffer and  
127 samples are boiled for 5 min.

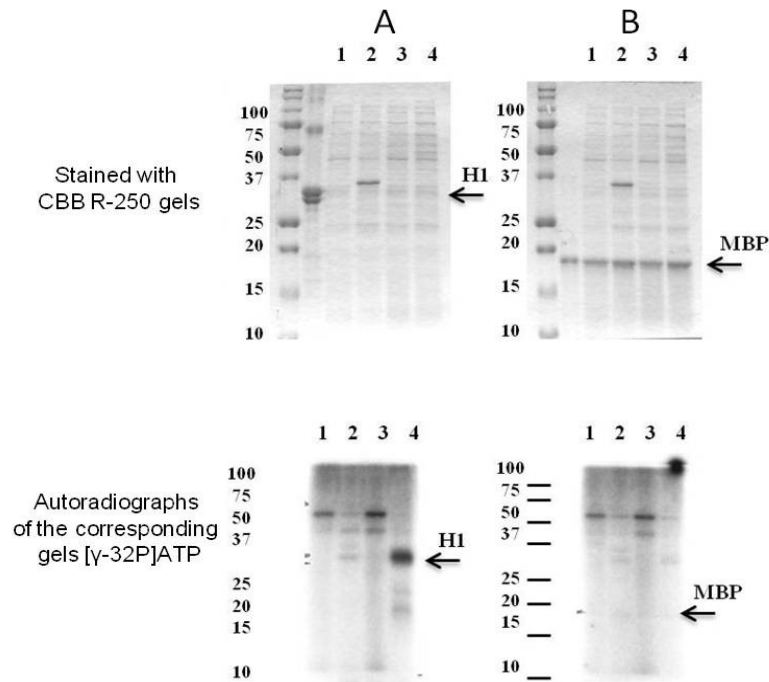
128 3. Phosphorylated products are resolved on SDS-polyacrylamide gels. The gels are then stained  
129 with CBB R250 as mentioned above, dried, and subjected to autoradiography. Exposition of dried  
130 gels with the X-Ray film varies from few hours at room temperature to few days at -70 °C and  
131 depends on the quantity and the activity of the recombinant protein kinase.

132 4. If the recombinant protein kinase is enzymatically active it can demonstrate either  
133 autophosphorylation or phosphorylation of the substrate exogenously provided. So, it is possible to  
134 make a decision about substrate specificity of the recombinant protein kinase examined.  
135 According to the results obtained we can conclude that at the phosphorylation condition  
136 described here the examined recombinant protein kinase is capable to phosphorylate histone H1.  
137 Since at the autoradiograph there is no a signal corresponding to apparent molecular weight  
138 of the recombinant protein kinase it does not undergo autophosphorylation.

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### 140 **Representative data**

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142

143 **Figure 1. The substrate specificity of recombinant SpkE.** Coomassie R-250 staining (at the top) and  
 144 autoradiographs of the corresponding gels (at the bottom). As potential substrates were examined (A)  
 145 histone H1 and (B) MBP. Soluble proteins were isolated from *E. coli* cells with pET41a(+) (1) before and  
 146 (2) after induction with IPTG and *E. coli* cells with pET41a(+) carrying *Synechocystis* spkE (3) before and  
 147 (4) after induction with IPTG. About 10 µg of the *E. coli* cells soluble protein fraction was mixed in 25 µl  
 148 with 5 µg either H1 or MBP in the presence of 1.5 mCi of [ $\gamma$ - $^{32}$ P]ATP. The phosphorylation reaction was  
 149 carried out for 15 min at 30 °C. As demonstrated above only histone H1 is phosphorylated by SpkE.  
 150 [modified from Zorina *et al.* (2014)]

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152 **Recipes**

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- 154 1. Homogenization buffer  
 155 50 mM Tris-HCl (pH 7.6)  
 156 10 mM MgCl<sub>2</sub>  
 157 2 mM EDTA  
 158 1 mM DTT  
 159 1 mM EGTA  
 160 2 mM Na<sub>3</sub>VO<sub>4</sub>  
 161 10 mM NaF  
 162 50 mM β-glycerophosphate  
 163 250 mM sucrose

- 164 1 mM PMSF  
 165 1 mM benzamidine hydrochloride  
 166 0.5 mM 6-aminocaproic acid  
 167 2. Reaction mixture (total volume 25  $\mu$ l)  
 168 20 mM MES-KOH (pH 6.5)  
 169 10 mM MgCl<sub>2</sub>  
 170 10  $\mu$ M ATP (unlabelled)  
 171 0.25 mg/ml MBP, or 0.5 mg/ml H1, or 1 mg/ml BSA  
 172 2  $\mu$ Ci [ $\gamma$ -<sup>32</sup>P]ATP  
 173 Protein kinase  
 174 ddH<sub>2</sub>O

175

### 176 **Acknowledgments**

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181

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