

Transmission Electron Microscopy for Tobacco Chloroplast Ultrastructure

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[Abstract] The chloroplast is the site of photosynthesis that enabled and sustains aerobic life on Earth. Chloroplasts are relatively large organelles with a diameter of ~5 µm and width of ~2.5 µm, and so can be readily analysed by electron microscopy. Each chloroplast is enclosed by two envelope membranes, which encompass an aqueous matrix, the stroma and the thylakoids. Components of stroma include starch granules and plastoglobuli, which can be observed by electron microscopy. And the thylakoids consist of stromal thylakoid, granal thylakoid and as well as granum (a stack of thylakoids). These structure components are quite sensitive to developmental changes and environmental variations, such as drought, salinity, cold, high temperature and others. Transmission electron microscopy (TEM) is a powerful technique for monitoring the effects of various changing parameters or treatments on the development and differentiation of these important organelles. Here we describe a reliable method for the analysis of plastid ultrastructure in tobacco plant by TEM.

Materials and Reagents

1. Tobacco (*Nicotiana tabacum*) plants (about 6-week grew on MS agar plates, 3-week grew in 1/4 Hoagland solution, and 2~3 weeks grew on soil)
2. Glutaraldehyde (Wako Pure Chemical Industries, catalog number: 071-01931)
3. Osmium tetroxide (Nisshin EM Corporation, catalog number: 300)
4. Ethanol (Wako Pure Chemical Industries, catalog number: 057-00456)
5. Propylene oxide (Wako Pure Chemical Industries, catalog number: 165-05026)
6. Quetol 812 set (Nisshin EM Corporation, catalog number: 340)
7. Uranyl acetate (Wako Pure Chemical Industries, catalog number: 6159-44-0)
8. 3% (w/v) lead citrate (Wako Pure Chemical Industries, catalog number: 121-01722)
9. Dodecenyl succinic anhydride (DDSA)
10. Methyl nadic anhydride (MNA)
11. DMP
12. 0.1 M phosphate buffered saline (see Recipes)
13. 1% osmium tetroxide (see Recipes)
14. Quetol-821 resin (see Recipes)
15. Hoagland solution (see Recipes)

Equipment

1. Razor blade
2. Lens tissue
3. Scissors
4. Tweezers
5. Needle and thread
6. Glass tube
7. Vacuum equipment
8. Balance
9. Petri dish
10. Oven at 60 °C
11. Plastic flat embedding mold (catalog number: 70900)
12. Beaker
13. Ultramicrotome (RMC, model: MT-7000)
14. Copper grid
15. Transmission electron microscope (JEOL, model: JEM-100CX II)

Procedure

1. Cut and trim tobacco leaf with a razor blade to 2 mm x 4 mm size, cover the leaf with lens tissue and tie it with needle and spread which can protect the sample and facilitate the fixation and the following steps (because the sample is easily floating and covered with lens tissue will facilitate sinking during fixation), then put it into 2% glutaraldehyde in a glass tube.
2. Vacuum the glutaraldehyde tube by vacuum equipment until no air bubble is coming out, and exhaust slowly to let the fix solution go into leaf tissue.
3. Fix in 2% glutaraldehyde for 2 h at room temperature.
4. Wash with 0.1 M PBS for 4~5 times and each time for 20~30 min.
5. Fix in 1% osmium tetroxide for 2 h.
6. Wash with distilled water for 30 min during which change the distilled water 5~8 times.
7. Add 50% ethanol and incubate for 10 min.
8. Discard 50% ethanol, add 70% ethanol and seal with Parafilm, keep overnight.
9. Discard 70% ethanol, add 90% ethanol for 10 min.
10. Discard 90% ethanol, wash with 100% ethanol for 3 times and each time for 10 min.
11. Add propylene oxide for 2 times and each time for 5 min.
12. Put the leaf sample into propylene oxide: Quetol 812 resin = 1:1 (v/v) and covered with aluminum foil overnight.
13. Change to Quetol 812 resin and keep for 24 h.

16. Embed the leaf sample horizontally with Quetol 812 resin on the Plastic flat embedding mold, and put into 60 °C for 48 h.
14. Cut to thin sections (70~100 nm) with a diamond knife on an ultramicrotome, and the thin sections are gathered on a copper grid which is specially used for transmission electron microscope observation (Figure 1).

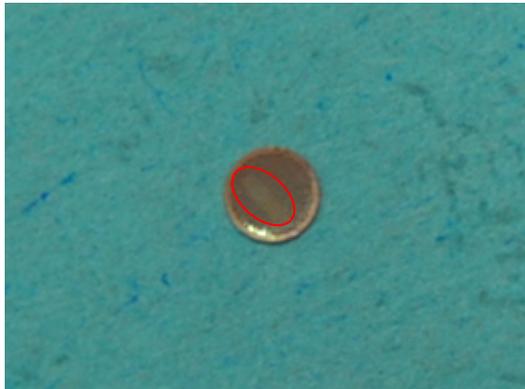


Figure 1. A picture of copper grid with sample sections. The red circle indicated the sample sections.

15. Stain with 2% (w/v) uranyl acetate for 40 min by putting copper grid with the sample (the sample side is downward) in staining solution and keep for 40 min, the following steps 16-18 are same.
16. Wash with distilled water in a beaker for 3 times and each time for 10 min gently.
17. Stain with 3% (w/v) lead citrate for 2~3 min.
18. Wash with distilled water in a beaker for 3 times and each time for 10 min gently.
19. Put on filter paper to make it dry and keep into the case.
20. Observe the samples on a transmission electron microscope at 80 kv, and take photos. The sample image of chloroplast can be found in Figure 6 in the Wang *et al.* (2014).

Notes

1. In step 2, be sure to exhaust slowly to let the fix solution go into the sample tissue.
2. During ethanol stepcoppers, keep a small amount of former solution before add new solution, this can avoid the dry of tissue edge.
3. When prepare the resin, be sure to mix well before adding DMP-30 accelerator. And the ratio of each reagent is important, but the total amount can be adjusted.
4. In steps 15 and 17, stain with uranyl acetate, drop a few drops on a Parafilm in a petridish, and put the copper disc on the drop.

Recipes

1. 0.2 M phosphate buffered saline (pH 7.0)

Solution A: 0.2 M Na₂HPO₄, Na₂HPO₄·12H₂O 7.164 g in 100 ml distilled water

Solution B: 0.2 M NaH₂PO₄, NaH₂PO₄·2H₂O 3.121 g in 100 ml distilled water

Put 61 ml solution A and 39 ml solution B together and mix well to make 0.2 M phosphate buffered saline at pH 7.0
2. 1% osmium tetroxide

4% osmium tetroxide: Distilled water: 0.2 M PBS= 1:1:2
3. Quetol-821 resin (in the case of prepare 102 g)

Quetol-812	48 g
DDSA	19 g
MNA	33 g
DMP	302 g
4. Hoagland solution

Note: Recipe can be found in Epstein (1972).

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