

Perls Staining for Histochemical Detection of Iron in Plant Samples

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[Abstract] Visualization of iron (Fe) localization in plants has greatly enhanced our understanding of plant Fe homeostasis. One of the relatively simple and yet powerful techniques is the classical Perls blue stain (Perls, 1867). The technique is based on the conversion of ferrocyanide to insoluble crystals of Prussian blue in the presence of Fe^{3+} under acidic conditions. It has been extensively used in animal and human histology (Meguro *et al.*, 2007) and has recently gained popularity in plant research. For specific purposes, Fe signals may be additionally enhanced in the 3,3'-diaminobenzidine tetrahydrochloride (DAB) intensification procedure (Meguro *et al.*, 2007). It has been demonstrated that this intensification results in the detection of both Fe^{2+} and Fe^{3+} ions (Roschztardt *et al.*, 2009). The method has been successfully applied at the whole plant, organ and subcellular levels, both with (Roschztardt *et al.*, 2011; Schuler *et al.*, 2012; Roschztardt *et al.*, 2013; Ivanov *et al.*, 2014) and without intensification (Stacey *et al.*, 2008; Long *et al.*, 2010).

Here, we present a full Perls staining and DAB intensification protocol, the way it is performed in our lab (Ivanov *et al.*, 2014).

Materials and Reagents

1. 3,3'-diaminobenzidine tetrahydrochloride (DAB) (Sigma-Aldrich, catalog number: 32750)
2. Chloroform (CHCl_3)
3. Cobalt (II) chloride (CoCl_2)
4. Ethanol ($\text{CH}_3\text{CH}_2\text{OH}$)
5. Glacial (water-free) acetic acid (CH_3COOH)
6. Hydrogen peroxide (H_2O_2) (30%)
7. Hydrochloric acid (HCl) (37%)
8. Methanol (CH_3OH)
9. Di-sodium hydrogenphosphate (Na_2HPO_4)
10. Sodium di-hydrogenphosphate (NaH_2PO_4)
11. Sodium azide (NaN_3)
12. Potassium ferrocyanide ($\text{K}_4[\text{Fe}(\text{Cn})_6]$)
13. Fixing solution (see Recipes)

14. Staining solution (see Recipes)
15. 0.1 M phosphate buffer (see Recipes)
16. 1% 3,3'-diaminobenzidine tetrahydrochloride (DAB) stock (see Recipes)
17. Preparation solution (see Recipes)
18. 1% CoCl_2 (see Recipes)
19. Intensification solution (see Recipes)

Equipment

1. Vacuum pump (any model capable of producing 500 mbar vacuum)
2. 1.5 ml tube
3. Standard incubator

Procedure

A. Perls stain

1. Fix plant material for 1-2 h under vacuum (500 mbar) in the fixing solution. Usually this can be done with 1 ml solution in a standard 1.5 ml tube. Larger samples will require recalculating the volumes. Vacuum application allows rapid penetration of the fixative. Samples need to be completely submerged. Using of vacuum desiccator is fully sufficient for this step. During this time, prepare the staining solution and pre-warm it at 37 °C in a standard incubator.
2. Remove the fixing solution.
3. Wash 3 times for 1-2 min each with distilled water.
4. Add the pre-warmed staining solution and incubate for 15 min to 1 h under vacuum (500 mbar). Again, the usual volume is 1 ml solution in a 1.5 ml tube.
5. Remove the staining solution.
6. Wash 3 times for 1-2 min each with distilled water. Gentle shaking might be applied.
7. Store in distilled water. It is advisable to analyze, or image, the samples within one week.
8. (Optional step) Dehydration: Incubate in an ethanol dilution series: 10%, 30%, 50%, 70%. Dehydration will intensify the blue stain. This step is not recommended if further intensification of the signal is planned. For samples with high iron content, such as roots or samples from metal hyperaccumulating plants, Perls stain alone may be fully sufficient for obtaining intensive signal. If the aim is a detailed view of tissue or cellular level iron detection the additional intensification should be performed (see below).

B. DAB intensification

9. Incubate the samples, prepared and washed in distilled water in step A6, for 1 h with the preparation solution. No vacuum is applied here.
10. Remove the solution and wash three times with 0.1 M Phosphate buffer.
11. Incubate the samples with Intensification solution at room temperature. Brown staining may appear as early as five min after the beginning of the incubation. In any case, the reaction should not continue for more than 30 min.
12. To stop the reaction, remove the Intensification solution and wash three times with distilled water.
13. Store samples in distilled water.

Notes

1. Incubation of plants in Fe-containing agar medium (usually around 50 μM Fe) will result in strong staining of the root apoplast. If the aim of the experiment is to observe whole-mount Fe distribution under standard Fe supply in inner parts of the root, such as the central cylinder, plants should be grown on soil.

Representative Data

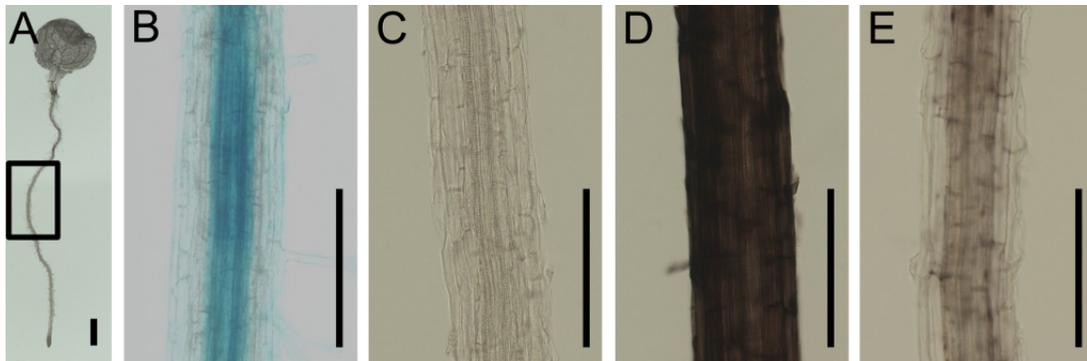


Figure 1. Visualization of Fe in *Arabidopsis* root by Perls staining and DAB intensification. A. A fixed 7 day-old *Arabidopsis* seedling. B. Perls-stained root. Signal is seen in the apoplast and central cylinder. C. DAB intensification applied without prior Perls staining. No specific signal can be seen. D. Perls-DAB staining on a root, grown on agar plate under sufficient Fe supply. Strong signal can be seen in the apoplast. E. Perls-DAB staining on a root, grown on agar plate under sufficient Fe supply and then transferred for 24 to Fe-deficient medium. Staining can be seen in the central cylinder. Low to no signal is present at the apoplast. The zone represented on the closeup images corresponds to the highlighted zone in A. Bars represent 1 mm.

Recipes

1. Fixing solution

Methanol: Chloroform: Glacial acetic acid (6:3:1)

The three can be pipetted in any order. The end solution should contain only one phase.

2. Staining solution

4% $K_4Fe(Cn)_6$ (4 g per 100 ml in distilled water) and 4% HCl (10.81 ml from a 37% HCl stock solution to a final volume of 100 ml) stock solutions are mixed in a 1:1 (v: v) proportion. It is advisable to store the two stock solutions in the dark at room temperature and prepare the mixture fresh before use.

3. 0.1 M phosphate buffer

Prepare 1 M stocks of Na_2HPO_4 and NaH_2PO_4 . For 50 ml 0.1 M buffer at pH 7.0, mix 2.89 ml of the Na_2HPO_4 and 2.12 ml of the NaH_2PO_4 . Add distilled water to a volume of 45 ml and measure the pH. If needed, adjust with sodium hydroxide (NaOH) or phosphoric acid (H_3PO_4). Then add distilled water to a final volume of 50 ml. pH value of the buffer is critical for the quality of the staining and may be varied, depending on the exact application. pH values under 7.0 will lead to reduction in staining intensity. pH over 7.6 may increase the background staining.

4. 1% 3,3'-diaminobenzidine tetrahydrochloride (DAB) stock

Several (10-15) microliters of 10 M (37%) HCl should be added per each milliliter of solution until the color becomes light brown. Shake for five minutes to help the complete dissolving of DAB. However, aggregates of undissolved DAB may remain. In this case, it is advisable to filter the solution through a 0.2 μ m filter. We have not tried to dissolve the remaining clumps by sonication.

5. Preparation solution

0.01 M NaN_3 and 0.3 % H_2O_2 in methanol

6. 1% $CoCl_2$

Stock solution needed for the preparation of the intensification solution

Can be stored for long periods at room temperature

7. Intensification solution

0.1 M Phosphate buffer (pH 7.0) containing 0.025% DAB, 0.005% H_2O_2 and 0.005% $CoCl_2$ Prepare fresh

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