

Escherichia coli* Infection of *Drosophila

Charles Tracy¹ and Helmut Krämer^{1, 2, *}

¹Department of Neuroscience, University of Texas Southwestern Medical Center, Dallas, USA;

²Department of Cell Biology, University of Texas Southwestern Medical Center, Dallas, USA

*For correspondence: helmut.kramer@utsouthwestern.edu

[Abstract] Following septic insults, healthy insects, just like vertebrates, mount a complex immune response to contain and destroy pathogens. The failure to efficiently clear bacterial infections in immunocompromised fly mutants leads to higher mortality rates which provide a powerful indicator for genes with important roles in innate immunity. The following protocol is designed to reproducibly inject a known amount of non-pathogenic *E. coli* into otherwise sterile flies and to measure the survival of flies after infection. The protocol can be easily adapted to different types of bacteria.

Keywords: *Drosophila*, Innate immunity, Bacterial infections, Tolerance, Survival

[Background] Classic infection experiments involve infecting *Drosophila* orally (Chakrabarti *et al.*, 2016) or with a needle dipped in a concentrated bacterial solution (Romeo and Lemaitre, 2008). Unlike these protocols, our experimental procedure allows us to determine the site of infection and precisely control the dose of bacteria injected into each fly. This provides homogeneity and reproducibility, and allows us to adapt bacterial load for different experiments (Akbar *et al.*, 2011 and 2016).

Materials and Reagents

1. 15 ml Falcon tubes (Corning, catalog number: 352196)
2. 1.5 ml Eppendorf tubes (USA Scientific, catalog number: 1615-5500)
3. Kimwipes
4. 26 G needle (BD, catalog number: 305111)
5. Spin columns (e.g., Thermo Fisher Scientific, Thermo Scientific™, catalog number: 69700)
6. Empty and clean *Drosophila* vials (Genesee Scientific, catalog number: 32-109)
7. Microloader tips (Eppendorf, catalog number: 930001007)
8. Vials (Genesee Scientific, catalog number: 32-109) with standard *Drosophila* food (lightly yeasted)
9. Microslide (Corning, catalog number: 2948-75X25)
10. 50 *Drosophila melanogaster* adult virgins (Romeo and Lemaitre, 2008), aged five to seven days post eclosion (see Note 1)
11. *E. coli*-DH5α containing any ampicillin-resistant plasmid (e.g., expressing GFP)
 - a. pEGFP (<https://www.addgene.org/vector-database/2485/>)
 - b. pET-GFP-C11 (<http://www.addgene.org/30183/>)

12. LB broth (Fisher Scientific, catalog number: BP1426-500)
13. Ampicillin (200 mg/ml) (Sigma-Aldrich, catalog number: A9518)
14. 70% ethanol–diluted from 100% ethanol (Pharmco-AAPER, catalog number: 111000200)
15. Mineral oil (Fisher Scientific, catalog number: O121-1)
16. Sodium chloride (NaCl) (Fisher Scientific, catalog number: S271)
17. Potassium chloride (KCl) (Sigma-Aldrich, catalog number: P9541)
18. Sodium phosphate dibasic heptahydrate ($\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$) (Sigma-Aldrich, catalog number: S9390)
19. Potassium phosphate monobasic (KH_2PO_4) (Fisher Scientific, catalog number: S397-500)
20. 1x PBS (see Recipes)

Equipment

1. 37 °C incubator with shaking (Eppendorf, New Brunswick™, model: Innova® 44)
2. Spectrophotometer (Molecular Devices, model: SpectraMax M2)
3. Borosilicate glass capillaries (WPI, catalog number: TW100-4)
4. Centrifuge capable of spinning Falcon tubes (Eppendorf, model: 5804 R)
5. Flaming/Brown Micropipette puller (Sutter Instrument, model: P-97)
6. Mini benchtop centrifuge (Fisher Scientific, model: Fisherbrand™ Standard Mini Centrifuge, catalog number: 05-090-100)
7. Pico-Injector (Nikon Instruments, catalog number: PLI-188)—requires nitrogen gas
Note: Should have foot pedal for injecting.
8. 25 °C incubator (BioCold Environment, model: BC49-IN)
9. Anesthetizing fly pad (Genesee Scientific, catalog number: 59-119)
10. Small brush
11. Dissecting microscope (Leica)

Software

1. Prism (GraphPad) software

Procedure

1. Preparation of *E. coli* suspension
 - a. Inoculate 50 ml LB broth with an *E. coli* colony from an LB agar plate under sterile conditions.
 - b. Add ampicillin (200 µg/ml final).
 - c. Incubate overnight at 37 °C with shaking. Use spectrophotometer to measure optical density at 600 nm (OD_{600}).
 - i. Dilute 500 µl aliquot of bacterial over-night suspension 5-fold with 2 ml of LB broth

- ii. Measure OD₆₀₀ of diluted bacteria with broth, and broth alone.
 - iii. Subtract OD₆₀₀ of fresh LB broth from bacteria with broth and calculate OD₆₀₀ of over-night culture.
 - d. Dilute over-night 50 ml bacterial culture to OD₆₀₀ = 0.1 with LB broth.
 - i. Spin 50 ml of bacterial suspension 4,000 × g for 10 min in Falcon tubes at room temperature.
 - ii. Decant LB broth.
 - iii. Re-suspend in 50 ml of 1x PBS by vortexing.
 - iv. Adjust with PBS to OD₆₀₀ = 0.1.
 - e. Heat-Kill bacteria (optional; see Note 2)
 - i. Aliquot 1 ml bacterial suspension into 1.5 ml Eppendorf tubes.
 - ii. Incubate at 65 °C for 20 min.
 - iii. Bacterial solution is now ready for injections.
2. Preparation of injection needles
- a. Place glass capillary in Micropipette puller.
 - b. Run program to create needles tapered like the needle pictured (Figure 1).
 - i. Ramp + 5, Pull = 90, Velocity = 70, Delay = 70

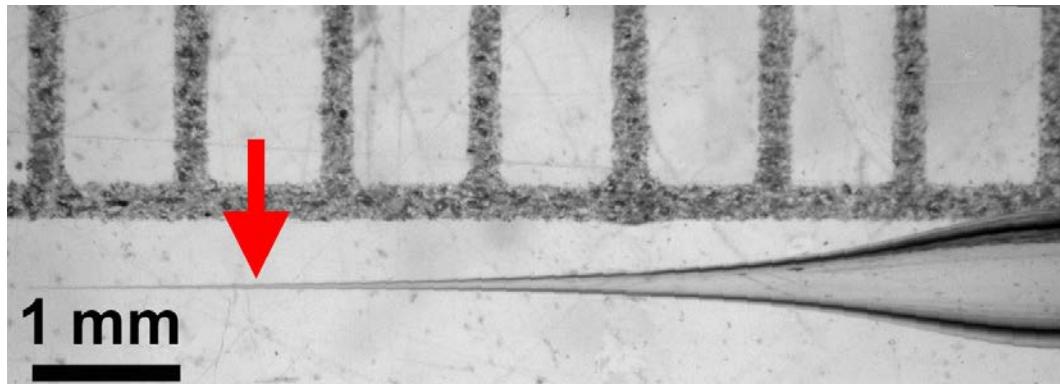


Figure 1. Tapered injection needle. Arrow indicates preferred place to break it.

- c. Stretch a Kimwipe taut over a Falcon tube and stab needle through the Kimwipe to break the very tip of the needle.
3. Fly sterilization
- a. Use a small needle (26 G) to poke three to four holes in the paper filter of the spin columns and place back into collection tube.
 - b. Add 25 anesthetized flies to one spin column (Figure 2).

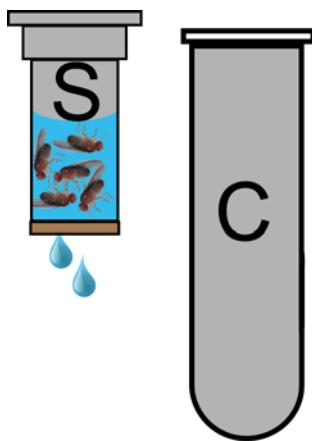


Figure 2. Spin column for Fly sterilization. 25 flies are loaded into a spin column (S) and covered with 70% ethanol. Spin columns are placed in a collection tube (C) to contain the flow through of wash solutions in the centrifuge during the quick spin. Flies are retained in the spin column by the paper filter at its bottom.

- c. Add 700 μ l 70% ethanol
 - d. Quick spin on mini benchtop centrifuge (~3-4 sec).
 - e. Remove spin column from collection tube and pour out flow through and place spin column back into collection tube.
 - f. Repeat ethanol wash steps 3c-3e.
 - g. Wash with 700 μ l sterile water.
 - h. Quickspin on mini benchtop centrifuge (~3-4 sec).
 - i. Repeat water rinse steps 3g and 3h.
 - j. Fold Kimwipe and place in empty fly vial.
 - k. Put flies into vial with Kimwipe, plug and place at 25 °C until flies have recovered.
4. Calibration of injection volume (see Note 3)
 - a. Load 1 μ l bacterial solution into the pulled needle using Microloader tips.
 - b. Attach needle to Pico-injector.
 - c. Adjust Pico-injector pressures, injection time to achieve 50 nl per injection.
 - i. Typical starting values for our set-up:
 - 1) Balance pressure = -0.2 psi—Should be neutral.
 - 2) Injection pressure = 15 psi.
 - 3) Injection time = 2 msec.
 - ii. For calibration: put a drop of mineral oil on a microslide.
 - iii. Inject bacterial solution into oil to determine nl per injection (a needle loaded with 1 μ l should yield 20 equal sized droplets of 50 nl each).
 - iv. Adjust injection time and pressure until desired amount is injected.
 - v. Optimal Pico-injector values will vary slightly between needles.
 5. Injection of sterilized flies (see Notes 4 and 5)

- a. Sterilize fly pad with 70% ethanol, allow to dry.
- b. Place sterilized flies on fly pad.
- c. Under dissecting microscope hold a fly in place by its legs with a small brush.
- d. Hold injecting rod and needle with other hand.
- e. Insert needle into the notum of the immobilized fly (Figure 3).
- f. Inject bacterial solution into fly using foot pedal—Audible beep will confirm.
- g. Repeat until all flies have been injected.



Figure 3. Injection of flies with bacterial suspension. Anesthetized flies are held with a brush as they are injected using a pulled glass needle (see Figure 1) filled with a bacterial suspension.

6. Incubate flies and monitor survival
 - a. Place ≤ 15 injected flies into vial containing fresh standard *Drosophila* media.

Vials should be lightly yeasted and contain no more than 1.5 ml of fly food. This minimizes the danger of flies getting stuck in yeast or between the side of the vial and the food as the food tends to dry at later days and separate from the vial.
 - b. Place vials with flies into 25 °C incubator and note how many flies are alive 2 h after injection.

Flies that do not recover from injections should be discarded from the dataset and subsequent analysis.
 - c. Count dead and living flies every day, once a day until all are dead. For bacteria more pathogenic than *E. coli*, shorter time intervals will be important.
 - d. Flip flies into fresh vials every 5 days.

Data analysis

Recorded data are entered into Prism (GraphPad) software which is used to display survival curves (Figure 4) and analyze statistical significance of differences in survival between genotypes. Log-Rank (Mantel-Cox) test was used to assess statistical significance of differences between survival curves (Figure 4). Alternatively, multiple Log-rank analysis software packages are available online at: <http://astatsa.com/> and <http://bioinf.wehi.edu.au/software/russell/logrank>. Both sides offer detailed information on how to enter data.

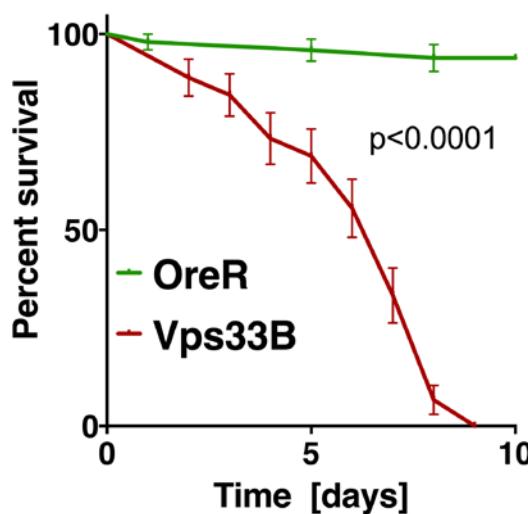


Figure 4. Example of statistical analysis of fly survival post bacterial infection. The diagram shows the fraction of surviving wild-type flies (OreR) or *Vps33B* mutants after injection with heat-killed *E. coli*, similar to experiments previously published (Akbar *et al.*, 2016). Statistical analysis was performed in Prism using the Log-rank (or Mantel-Cox) test to determine a *P* value smaller than 0.0001.

Notes

1. A wild-type *Drosophila* strain (Oregon R or Canton S) should be included as a negative control group for injections. Use sibling and/or parent flies as controls when appropriate. Three replicates of fifty flies per genotype are sufficient for proper statistical analysis.
2. It is important to note that heat inactivation can change the potential of bacteria to engage different innate immunity pathways as normally hidden epitopes may be exposed (Chung and Kocks, 2011).
3. Most difficulties in this experimental procedure are encountered during the needle calibration and injection steps. The balance pressure is the source of most problems. If the balance pressure is set too high, the result is a constant bleeding of the bacterial solution resulting in difficulty measuring injection droplets, emptying of the needle, and dripping bacterial solution

onto the fly. No liquid should escape the needle until the injection foot pedal has been pressed. Problems also arise if the balance pressure is too low. Low pressure will result in sucking up liquid that had just been injected resulting in little to no bacteria being injected (this should also be part of your calibration). Furthermore, once the needle is inserted into the fly, the negative pressure will pull fly tissues into the needle tip resulting in blockage that can often be seen through the microscope. This will prevent any further injections when the foot pedal has been pushed. Placing the needle tip into the bacterial solution and pressing the ‘clear’ button on the Pico-injector can usually solve this by high positive pressure clearing the needle. Once the blockage is cleared, leave the needle tip in the bacterial solution and hold the ‘fill’ button to refill the needle using negative pressure. Once the needle is calibrated it is also useful to inject a few droplets on or near a fly or paintbrush to get a good sense of the size of a 50 nl droplet. A 50 nl droplet is roughly the size of a fly eye.

4. Prior to commencing bacterial injections, it is important to have a series of experiments showing flies do not die from inflammation resulting from the needle injury or from the injection media. Mock injections should be performed on wild-type and experimental flies to determine if injury alone is sufficient to kill flies. Furthermore, control injections of sterile PBS should be done to ensure flies die from infection. These mock injections should be analyzed in the same manner as injections of bacteria as described in the data analysis section.
5. The experimental procedure described above has been optimized for DH5α *E. coli*. Each 50 nl injection should deliver around 2,000 CFU to each fly. If other bacterial species are to be used, bacterial loads should be optimized to ensure survival of wild-type flies when virulence allows.

Recipes

1. 1x PBS
137 mM NaCl
2.7 mM KCl
10 mM Na₂HPO₄
1.8 mM KH₂PO₄

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This protocol has been adapted and modified from our previously published work (Akbar *et al.*, 2016; Akbar *et al.*, 2011).

The authors declare no conflicts of interest.

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