

## Determination of Mannitol-2-dehydrogenase Activity from *Zobellia galactanivorans*

Agnès Groisillier\* and Thierry Tonon

Sorbonne Université, UPMC Univ Paris 06, CNRS, Integrative Biology of Marine Models, Station Biologique de Roscoff, Roscoff cedex, France

\*For correspondence: [groisill@sb-roscoff.fr](mailto:groisill@sb-roscoff.fr)

**[Abstract]** Mannitol is a polyol that occurs in a wide range of living organisms, where it fulfills different physiological roles. Several pathways have been described for the metabolism of mannitol by bacteria, including the phosphoenolpyruvate-dependent phosphotransferase system (PST) and a M2DH-based catabolic pathway. The latter involves two enzymes, a mannitol-2-dehydrogenase (EC 1.1.1.67) and a fructokinase (EC 2.7.1.4), and has been identified in different bacteria, e.g. the marine Bacteroidetes *Zobellia galactanivorans* (Zg) which had recently gained interest to study the degradation of macroalgal polysaccharides. This protocol describes the biochemical characterization of a recombinant mannitol-2-dehydrogenase (M2DH) of *Zobellia galactanivorans*. The ZgM2DH enzyme catalyzes the reversible conversion of mannitol to fructose using NAD<sup>+</sup> as a cofactor. ZgM2DH activity was assayed in both directions, i.e. fructose reduction and mannitol oxidation.

### **Reversible reaction:**



### **Materials and Reagents**

1. UV-Star® PS Microplate (96 Well) (Greiner Bio-One GmbH, catalog number: 655801)
2. Purified recombinant His-tagged ZgM2DH  
*Note: This protein was produced in Escherichia coli BL21 (DE3) containing the recombinant pFO4\_ZgM2DH vector, as described by Groisillier et al. (2010). This recombinant protein was purified by affinity chromatography using a HisPrep FF 16/10 column (GE Healthcare Dharmacon) onto an Äkta avant system (GE Healthcare Dharmacon). The complete purification protocol is described in details in Groisillier et al. (2015).*
3. Trizma® base (Sigma-Aldrich, catalog number: T1503)
4. MilliQ water
5. 4-morpholineethane-sulfonic acid (MES) (Sigma-Aldrich, catalog number: M2933)
6. Bis-Tris (Sigma-Aldrich, catalog number: B9754)
7. Examples of chemicals to be tested to assess substrate and co-factor specificity:
  - a. D(-)-fructose (Sigma-Aldrich, catalog number: F0127)
  - b. D(+)-glucose (Sigma-Aldrich, catalog number: G8270)

- c. D-(+)-mannose (Sigma-Aldrich, catalog number: M4625)
- d. D-(+)-galactose (Sigma-Aldrich, catalog number: G0750)
- e. D-(+)-xylose (Sigma-Aldrich, catalog number: X1500)
- f. D-mannitol (Sigma-Aldrich, catalog number: M9647)
- g. D-sorbitol (Sigma-Aldrich, catalog number: S1876)
- h. D-(+)-arabitol (Sigma-Aldrich, catalog number: A3381)
- i.  $\beta$ -Nicotinamide adenine dinucleotide hydrate ( $\beta$ -NAD) (Sigma-Aldrich, catalog number: N1636)
- j.  $\beta$ -Nicotinamide adenine dinucleotide phosphate hydrate ( $\beta$ -NADP) (Sigma-Aldrich, catalog number: N5755)
- k.  $\beta$ -Nicotinamide adenine dinucleotide, reduced disodium salt hydrate ( $\beta$ -NADH) (Sigma-Aldrich, catalog number: N8129)
- l.  $\beta$ -Nicotinamide adenine dinucleotide phosphate, reduced tetra (cyclohexylammonium) salt ( $\beta$ -NADPH) (Sigma-Aldrich, catalog number: N5130)
8. 1 M Tris-HCl (pH 7.5) (see Recipes)
9. 10 mM NADH (see Recipes)
10. 10 mM NAD<sup>+</sup> (see Recipes)

### **Equipment**

1. Safire2 UV spectrophotometer microplate reader (Tecan Trading AG)
2. NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific)

### **Software**

1. Hyper 32 ([hyper32.software.informer.com](http://hyper32.software.informer.com))
2. Microsoft Excel

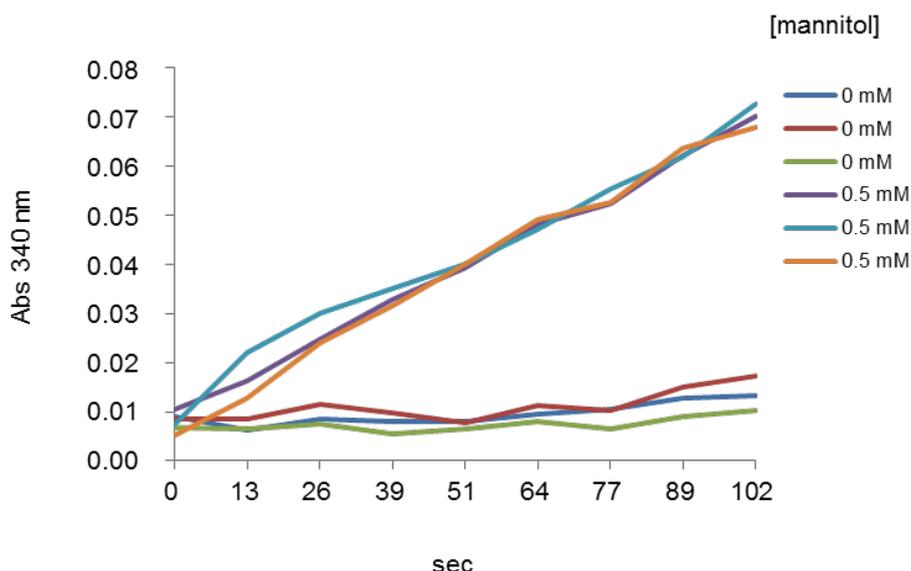
### **Procedure**

1. The standard fructose reduction reaction mixture contains 100 mM Tris-HCl (pH 6.5), 1 mM fructose, 0.2 mM NADH and 1 to 10  $\mu$ g of purified recombinant enzyme, in a final volume of volume 100  $\mu$ l. The standard mannitol oxidation reaction mixture contains 100 mM Tris-HCl (pH 8.5), 1 mM mannitol, 0.5 mM NAD<sup>+</sup> and 1 to 10  $\mu$ g of purified enzyme in 100  $\mu$ l. Blank corresponds to reaction mixture where substrate is substituted by MilliQ water (Table 1).

**Table 1. Composition of blank and reaction mixture for determination of ZgM2DH activity (fructose reduction)**

Stock solutions	µl added in blank	µl added in reaction mix (test)
Tris-HCl (pH 6.5) (1 M)	10	10
NADH (10 mM)	2	2
Fructose (25 mM)	0	4
ZgM2DH (1 µg/µl)	1	1
MilliQ water	87	83

- The continuous assay reaction is started by adding the substrate and the activity is monitored by following changes in absorbance at 340 nm, which corresponds to the decrease or production of NADH, in a Safire2 UV spectrophotometer microplate reader. The reaction is performed at 25 °C for up to 20 min. Only the early linear part of the curves is used to calculate activity (Figure 2).



**Figure 2. Early linear parts of curves representative absorbance (340 nm) monitored as a function of time (sec) in presence of mannitol.** The curves represent two series of triplicates at 0 and 0.5 mM of mannitol, respectively.

- Fructose reduction and mannitol oxidation activities, based on decrease or production of NADH respectively, are calculated using the formula:

$$[(\Delta A_{340\text{nm}} \text{ test} - \Delta A_{340\text{nm}} \text{ blank}) / (t * 6220 * 0.3 * 0.0001)]$$

Where

$\Delta A_{340\text{nm}}$  = variation of absorbance during the duration of incubation

t = duration of incubation (min)

6220 = extinction coefficient for NADH ( $\text{L mol}^{-1} \text{ cm}^{-1}$ )

0.3 = optical path (cm)

0.0001= assay volume (L)

One unit (U) of both activities (fructose reduction or mannitol oxidation) corresponds to 1  $\mu$ mol of NADH reduced or oxidized per min.

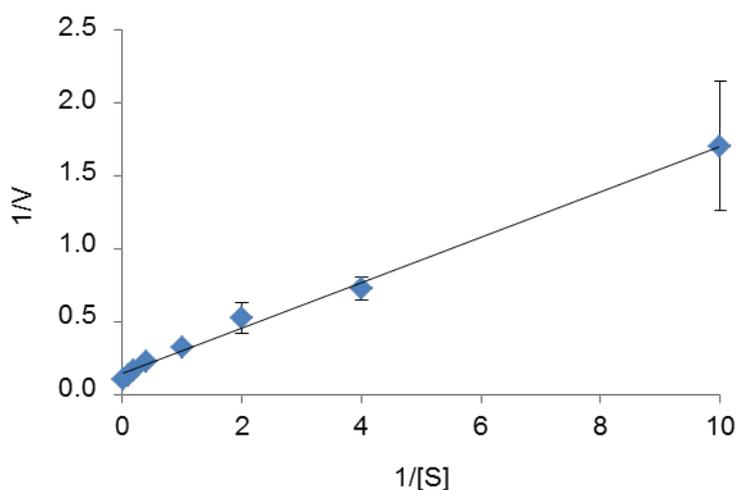
- To calculate specific activities, divide the value obtained in the equation above by the quantity of ZgM2DH proteins added in the reaction mixture. Perform three replicates for each assay, and determine the average  $\pm$  S. E. (*cf.* Notes) for these three replicates. This applies also to the experiments described below. As an example, typical value for specific activity of ZgM2DH was 3.95 U/mg in presence of 10 mM fructose.
- To determine substrate specificity, test ZgM2DH activity in presence of each substrate listed in the "Materials and Reagents" section, using concentration ranging from 1 mM to at least 100 mM.
- To determine the optimal pH, replace the 100 mM Tris-HCl (pH 8.5 or 6.5) buffer used in step 1 of procedure by other buffers prepared at different pHs. As an indication, it is possible to use:  
100 mM MES for pH 5.5, 6, 6.5  
100 mM Bis-Tris propane for pH 6.5, 7, 7.5, 8, 8.5, 9, 9.5  
100 mM Tris-HCl for pH 7, 7.5, 8, 8.5, 9
- To determine the optimal temperature, incubate the reaction mixtures used in step 1 at temperatures ranging, for instance, from 10 °C to 50 °C, with incremental of 5 or 10 °C.
- To assess the effects of various metal ions and chemicals on ZgM2DH activities, add metal ions or chemicals individually and at varying concentrations in the reaction mixture described in step 1. Examples are given in Table 2.

**Table 2. Metal ions and chemicals tested for potential influence on the ZgM2DH activities.** Mannitol, fructose, mannitol-1-phosphate and fructose-6-phosphate were used at high concentrations to assess potential inhibitory effects on enzymatic activity.

Compounds	Final concentration in the reaction mix (mM)
Ca <sup>2+</sup>	10
K <sup>+</sup>	10
Li <sup>+</sup>	10
NH <sub>4</sub> <sup>+</sup>	10
Mg <sup>2+</sup>	10
Mercaptoethanol	10
EDTA	10
Fructose	20
Fructose	50

Mannitol	20
Mannitol	50
Mannitol-1-phosphate	20
Fructose-6-phosphate	20
NaCl	100
NaCl	1,000

- To estimate the kinetic parameters of the enzyme for a selected substrate S, run individual enzyme reactions in presence of at least five different concentrations of this substrate and a fixed concentration of NAD(H). Determine the initial reaction rate for each reaction and plot  $1/V$  versus  $1/[S]$  to obtain a Lineweaver-Burk plot, from which  $K_m$  and  $V_m$  for S can be calculated (Figure 3).



**Figure 3. Lineweaver-Burk plot used to determine the  $K_m$  (1.12 mM) and  $V_m$  (7.18 U/mg) of ZgM2DH for mannitol.** [S] is the mannitol concentration (in mM) and V is the reaction rate (in U/mg of protein).

### Notes

- Before determining  $K_m$  and  $V_m$  for a given substrate S, be sure that cofactor [NAD(H)] is in excess, *i.e.*, that V does not increase with increasing quantities of cofactor in the reaction mixture; in the same vein, before determining  $K_m$  and  $V_m$  for the cofactor, be sure that S is in excess, *i.e.*, that V does not increase with increasing quantities of S in the reaction mixture. In theory, saturating concentration is equivalent to 100  $K_m$ , but 10  $K_m$  is usually sufficient (Bisswanger, 2014). It is then necessary to adjust the ZgM2DH concentration and the incubation time to obtain a linear change of absorbance at 340 nm, *i.e.*, a linear change of NADH.
- S. E. corresponds to standard error calculated in Excel.

## **Recipes**

1. 1 M Tris-HCl (pH 7.5)  
Stored at room temperature
2. 10 mM NADH  
Prepared fresh in water on the day of use
3. 10 mM NAD<sup>+</sup>  
Prepared fresh in MilliQ water on the week of use and stored at -20 °C

## **Acknowledgments**

This protocol was performed by Groisillier *et al.* (2015). This work was supported by the French National Research Agency via the investment expenditure program IDEALG (ANR-1 0-BTBR-02). We also acknowledge funding from the Émergence-UPMC-2011 research program.

## **References**

1. Bisswanger, H. (2014). [Enzymes assays](#). *Perspective in Sciences* 1: 41-55.
2. Groisillier, A., Herve, C., Jedy, A., Rebuffet, E., Pluchon, P. F., Chevlot, Y., Flament, D., Geslin, C., Morgado, I. M., Power, D., Branno, M., Moreau, H., Michel, G., Boyen, C. and Czjzek, M. (2010). [MARINE-EXPRESS: taking advantage of high throughput cloning and expression strategies for the post-genomic analysis of marine organisms](#). *Microb Cell Fact* 9: 45.
3. Groisillier, A., Labourel, A., Michel, G. and Tonon, T. (2015). [The mannitol utilization system of the marine bacterium \*Zobellia galactanivorans\*](#). *Appl Environ Microbiol* 81(5): 1799-1812.