

## Isolation of Heterocysts from *Anabaena* sp. PCC 7120

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**[Abstract]** During combined nitrogen step-down, filaments of cyanobacterium *Anabaena* sp. PCC 7120 differentiate about 5-10% of vegetative photosynthetic cells into heterocysts, the specialized cells for N<sub>2</sub> fixation (Walk, 1996). Heterocysts have a thick cell wall reducing permeation of O<sub>2</sub> and consist of two additional layers composed of glycolipids and polysaccharides. The difference in structure and composition of the cell wall between heterocysts and vegetative cells allows separation and isolation of heterocyst. Heterocysts isolated by this protocol can be subjected to protein analysis and activity measurements, which do not require strict anaerobic conditions.

### **Materials and Reagents**

1. *Anabaena* sp. PCC 7120
2. BG11<sub>0</sub> growth medium-BG11 medium without addition of combined nitrogen (Rippka *et al.*, 1979)
3. 1 M 2-[[1,3-dihydroxy-2-(hydroxymethyl)propan-2-yl]amino]ethanesulfonic acid-KOH (TES) (pH 8.2)
4. 1 M 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid-NaOH (Hepes) (pH 7.2)
5. 1 M NaCl
6. 1 M Na-EDTA
7. Sucrose (Merck KGaA, catalog number: K45136551417)
8. Lysozyme (N-acetyl muramide glycanhydrolase) (Roche Diagnostics, catalog number: 10837059001)
9. Extraction buffer (see Recipes)

### **Equipment**

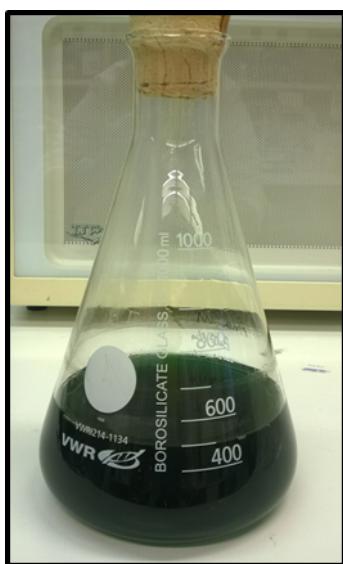
1. 50 ml Falcon tubes
2. 500 ml centrifuge vials (optional)
3. Spectrophotometer (e.g., PerkinElmer, model: Lambda 25)
4. Sorvall GS-3 rotor centrifuge for 500 ml vials (optional)
5. SL 16R centrifuge (6 x 50 ml) (e.g., Thermo Fisher Scientific)
6. Vortex
7. Sonicator (e.g., Labsonic U, B. Braun Melsungen AG)

8. Orbital shaker (e.g., Infors HT, Labotron)
9. Incubator (e.g., Certomat, B. Braun Melsungen AG)
10. Bright-field microscope (40x magnification)
11. Water bath or thermostat at 37 °C for Chlorophyll measurements

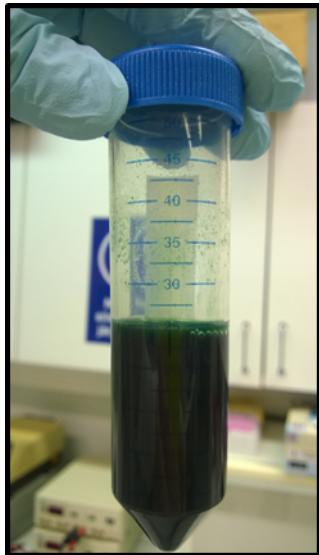
## **Procedure**

1. It is important to cultivate *Anabaena* sp. PCC 7120 in BG11<sub>0</sub> medium without combined nitrogen. In our laboratory, cultures are cultivated in BG11<sub>0</sub> supplemented with 10 mM TES-KOH (pH 8.2) at 30 °C under continuous light intensity of 50 µmol photons m<sup>-2</sup> s<sup>-1</sup> (PAR) and under agitation (about 120 RPM) to assure continuous gas exchange and kept at high CO<sub>2</sub> (air enriched with 3% CO<sub>2</sub>).
2. Harvest filaments during the logarithmic growth phase (OD<sub>750</sub> = 1). To obtain 10-30 ml of purified heterocysts at the Chlorophyll (Chl) a concentration of 15-20 µg/ml use about 300-400 ml of cyanobacterial culture.

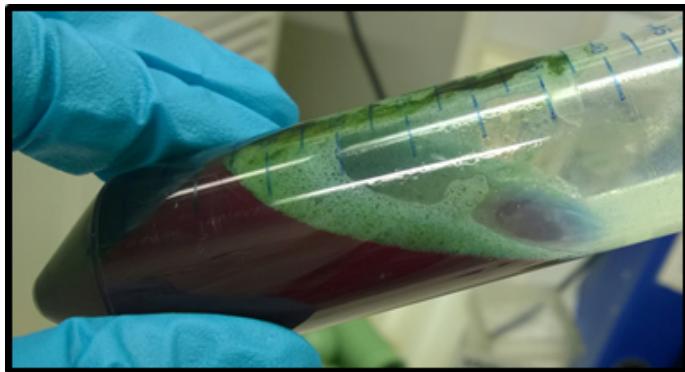
*Note: Chl a quantification, as described in Meeks and Castenholz (1971), is based on the extinction coefficient of 78,74 L/g/cm for Chl a in 90% methanol.*



3. Centrifuge filaments at 5,000 × g at room temperature for 5-10 min, discard the supernatant. If available, use a centrifuge with the rotor fitting 500 ml tubes or distribute the culture into 50 ml Falcon tubes and perform several rounds of centrifugations. In the end of centrifugation collect all cells in one 50 ml falcon tube and discard as much of growth medium as possible.
4. Keep the pellet in falcon tube and add extraction buffer till about 30 ml, vortex thoroughly.



5. Add 30 mg of fresh (!) lysozyme into the falcon tube, incubate for 1 h at 37 °C with continuous mixing on orbital shaker (200 RPM).
6. Sonicate at 40 W for 1 min in water bath filled with ice-cold water to avoid excessive heating. Lysozyme treatment and sonication will digest vegetative cells, whereas heterocysts will remain intact. After this step all procedures should be performed at 4 °C and on ice.
7. Centrifuge cell suspension at 1,000  $\times g$  for 5 min (4 °C), discard a supernatant (it can be used as a crude protein extract from vegetative cells).



8. Resuspend the pellet in 10-15 ml of ice-cold extraction buffer, vortex thoroughly, centrifuge at 250  $\times g$  for 3 min (4 °C) and discard the supernatant. Repeat washing minimum 3 times or until the supernatant is not blue; the residual pellet containing heterocysts should be light-green.

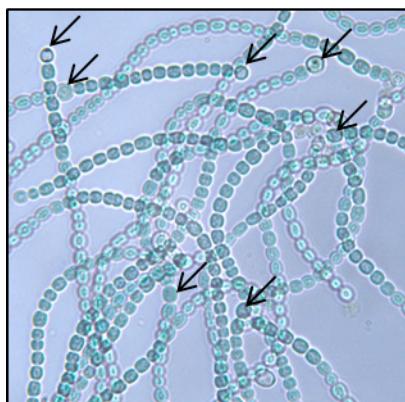


Filaments

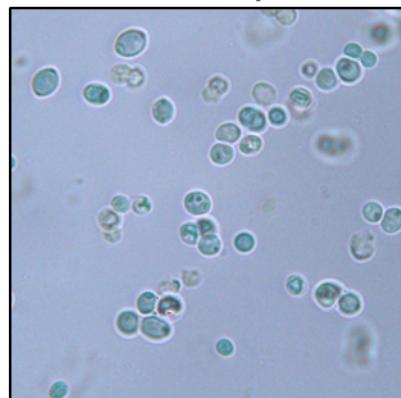
Heterocysts

9. Check purity of heterocysts by microscope. Take small amount of pellet on a glass slide (optionally resuspend in small amount of extraction buffer) and visualize cells under a microscope (40x magnification). The pellet must be mostly formed by aggregates of intact roundish pale-green heterocysts, in some cells polar granules of cyanophycin might be visible. Alcian blue staining might also be used to confirm the absence of vegetative cells in preparation (Maldener *et al.*, 2003).

Filaments



Heterocysts



*Note: Under the microscope vegetative cells and heterocysts can be easily distinguish by the shape of the cells and by the color. Due to the features of the camera, in this photo the color of cells is modified. Small arrows point to the heterocysts in filaments.*

*Note: If the pellet still contains filaments consisting of several cells, then lysozyme treatment was not successful. In this case repeat lysozyme treatment with a fresh enzyme and continue with sonication.*

10. Freeze the pellet in liquid nitrogen and store at -80 °C to use later for protein isolation or resuspend in extraction buffer to reach the desirable concentration of Chl a for immediate activity measurements. After freezing heterocysts do not retain the activity.

## Recipes

### 1. Extraction buffer

50 mM HEPES-NaOH (pH 7.2)

0.4 M sucrose

10 mM NaCl

10 mM EDTA

## Acknowledgments

The protocol was adopted from Ermakova *et al.* (2014), Fay (1980), Razquin *et al.* (1996), and Ow *et al.* (2009). The work was supported by the Academy of Finland (projects 271832) and Kone Foundation.

## References

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