

## Lysosomal Amino Acid Efflux Assay

Chunlei Cang\* and Dejian Ren

Department of Biology, University of Pennsylvania, Philadelphia, USA

\*For correspondence: [ccang@sas.upenn.edu](mailto:ccang@sas.upenn.edu)

**[Abstract]** As the cellular “recycling” organelle, lysosomes break down proteins into amino acids, which are then transported into cytosol for reuse by various amino acid transporters. The transport rate of an amino acid is presumably regulated by cellular conditions such as organelle pH, membrane potential and metabolic states. Because of their intracellular localization and the relative inaccessibility, lysosomal amino acid transporters have been studied largely via indirect measurements. Using lysosome purification and <sup>14</sup>C-labeled amino acids, this protocol provides a method to measure the efficiency of specific amino acid transporters on lysosomes.

### Materials and Reagents

1. Mouse liver
2. <sup>14</sup>C-labeled amino acids (PerkinElmer) (select the amino acid of your interest)
3. 3 N methanolic HCl (Sigma-Aldrich, catalog number: 33051)
4. Anhydrous methanol (Sigma-Aldrich, catalog number: 322415)
5. Sucrose
6. HEPES
7. MOPS
8. Na<sub>2</sub>EDTA
9. KCl
10. MgCl<sub>2</sub>
11. ATP-Mg
12. Phosphate buffered saline (PBS)
13. Scintillation cocktail (Research Products International, catalog number: 111175)
14. Protein assay kit (Bio-Rad Laboratories, catalog number: 500-0112)
15. Homogenization buffer (HB) (see Recipes)
16. High sucrose buffer (see Recipes)
17. Uptake buffer (see Recipes)
18. Efflux buffer (see Recipes)

## **Equipment**

1. Nitrogen gas tank (with regulator and connecting tube)
2. 25-ml glass conical flask
3. 1.5 ml and 15 ml tubes
4. 30 G needles
5. Dounce homogenizer
6. High speed centrifuge (up to 25,000 x g) with temperature control
7. Glassfiber filters (Grade GF/F) (Whatman, catalog number: 1825-025)
8. Vacuum filtration manifold (EMD Millipore, model: 1225)
9. Liquid scintillation counter (Beckman Coulter, model: LS6500)
10. Personal protective equipment for working with radioactive materials

## **Procedure**

*Note: This protocol involves  $^{14}\text{C}$ -labeled amino acids. The uses of radioactive isotopes and the necessary safety precautions to be taken should follow institutional regulations.*

### A. Preparation of amino acid methyl esters

1. Transfer 12.5  $\mu\text{Ci}$   $^{14}\text{C}$ -labeled amino acid into a 25-ml glass conical flask and let it dry under stream of nitrogen.

*Note: The drying time depends on the speed of nitrogen flow and is about 10 min.*

2. Add 3 ml 3 N methanolic HCl into the flask and incubate for 24 h at room temperature. This will transform amino acid into amino acid methyl ester.

*Note: Seal the opening of the flask with parafilm to prevent evaporation of methanolic HCl. The transformation efficiency can be measured with chromatography and high voltage electrophoresis (Reeves, 1979; Steinherz et al., 1982).*

3. Dry the resulted amino acid methyl ester under stream of nitrogen. Add 1 ml anhydrous methanol to dissolve the pellet and dry it again under stream of nitrogen.

*Note: The drying time depends on the speed of nitrogen flow.*

4. Add 500  $\mu\text{l}$  anhydrous methanol to dissolve the methyl ester. Bring down the volume to 250  $\mu\text{l}$  through evaporation under stream of nitrogen.

5. Shortly before efflux assay, transfer appropriate amount into a 1.5 ml tube. Dry under stream of nitrogen. Dissolve in uptake buffer (final concentration 0.005  $\mu\text{Ci}/\mu\text{l}$ ).

*Note: Amino acid methyl ester is not stable in aqueous solution.*

### B. Crude lysosome preparation

1. To prepare crude lysosome preparation from mouse liver, follow animal use protocols and dissect out the liver after the animal is euthanized. Rinse in ice-cold HB.  
*Note: Liver from one adult mouse is enough for one assay. If you need to test multiple amino acids, calculate the total protein concentration you can get from one mouse liver and scale up the animal number.*
2. Transfer the liver into a Dounce homogenizer, add 8 ml ice-cold HB.
3. Homogenize the liver on ice to break liver cells (20 strokes).
4. Centrifuge the homogenate at 1,000 x g for 10 min at 4 °C; transfer the supernatant into a new centrifuge tube and keep on ice.
5. Homogenize the pellet again in 7 ml HB (15 strokes); centrifuge at 1,000 x g for 10 min at 4 °C.
6. Combine the supernatant from steps B4-5; centrifuge at 20,000 x g for 20 min at 4 °C.
7. Resuspend the pellet in 10 ml 250 mM KCl (pH 7.4, buffered by 1 mM MOPS) and centrifuge at 1,600 x g for 10 min at 4 °C. Transfer the supernatant to a fresh tube. This step pellets most of the mitochondria.
8. Dilute the supernatant by 2.5x volume high sucrose buffer and centrifuge at 25,000 x g for 10 min to spin down the crude lysosomes.
9. Resuspend the crude lysosome pellet in 200 µl uptake buffer. Centrifuge at 600 x g for 2 min.
10. Transfer the supernatant to a new tube and keep it on ice. Resuspend the pellet in 100 µl uptake buffer by passing through a 30 G needle for 30 times. Combine it with the supernatant. This will be the crude lysosome sample.
11. Use a small aliquot of crude lysosome to measure total protein concentration with protein assay. Adjust the concentration to 2 mg protein-equivalent amount of lysosomes/ml. Keep the rest of the preparation on ice and do the efflux assay within 12 h. Longer storage is not recommended.

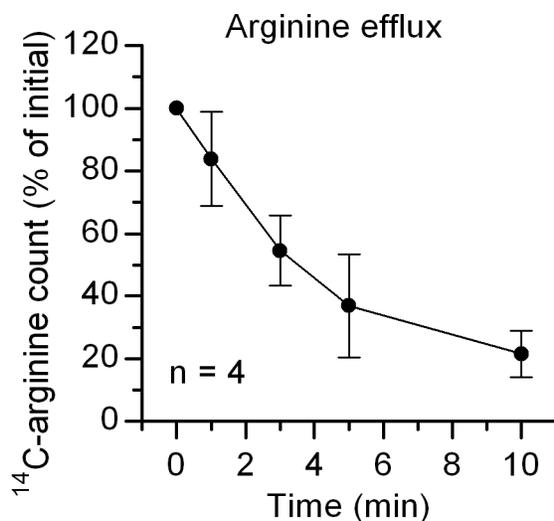
*Note: If needed, lysosomes can be further purified from the above preparation using, for example, OptiPrep density gradient centrifugation (see References 4 and 5).*

#### C. Amino acid efflux assay

1. For each reaction, dispense 50 µl aliquot (containing 100 µg of protein-equivalent amount of lysosomes) into a 1.5 ml tube.
2. To initiate amino acid ester loading, add 50 µl of amino acid methyl ester (0.25 µCi, in uptake buffer) into each lysosome aliquot tube. Gently mix by pipetting. Incubate at 37 °C for 30 min.

*Note: The optimum loading time and temperature need to be experimentally determined for the amino acid and the type of lysosomes used.*

3. During the loading time, assemble the Whatman filters (GF/F) to vacuum filtration manifold. Rinse the filters with 5 ml PBS. Adjust the filtering speed to ~10 ml/min.
4. At the end of loading, dilute the loading reaction 50x by transferring the 100  $\mu$ l loading reaction into a 15 ml tube containing 5 ml pre-warmed (37 °C) efflux buffer; mix well and incubate at 37 °C. This will start the efflux.
5. At the end of each efflux time point, combine a tube of efflux reaction (containing 5.1 ml) with 5 ml ice-cold PBS. Filter immediately through Whatman filter under vacuum; wash twice with 5 ml ice-cold PBS.
6. For background count, incubate lysosomes and amino acid methyl ester separately (without loading). Mix and filter immediately after incubation.
7. Soak the filter in scintillation cocktail overnight. Count using a scintillation counter.
8. Subtract background from the isotope counts. Normalize the background-subtracted counts to that obtained at efflux time point 0. The normalized count at each time point indicates the portion of amino acid remaining in the lysosome and reflects the activity of the amino acid transporter (see Figure 1 for an example).



**Figure 1. A representative amino acid efflux result using mouse crude lysosomes and <sup>14</sup>C-labeled arginine.** Data are presented as mean  $\pm$  SEM.

### Recipes

1. Homogenization buffer (HB)
  - 250 mM sucrose
  - 1 mM Na<sub>2</sub>EDTA
  - 10 mM HEPES (pH 7.0)

2. High sucrose buffer
  - 450 mM sucrose
  - 0.5 mM Na<sub>2</sub>EDTA
  - 5 mM HEPES (pH 7.2)
3. Uptake buffer
  - 250 mM sucrose
  - 5 mM MgCl<sub>2</sub>
  - 20 mM HEPES (pH 7.0)
4. Efflux buffer
  - 250 mM sucrose
  - 5 mM MgCl<sub>2</sub>
  - 20 mM HEPES
  - 2 mM ATP-Mg (pH 7.0)

### **Acknowledgments**

This protocol was adapted from previously published papers (Cang *et al.*, 2013; Reeves, 1979; Steinherz *et al.*, 1982; Graham *et al.*, 1982) and the technical bulletin of the lysosome isolation kit from Sigma-Aldrich (Reference 5). We thank Dr. Yandong Zhou and Dr. Youngjun Seo for discussion and conducting the experiments, and Dr. Denia Ramirez-Montealegre for advice on the protocol. The development of this protocol was supported, in part, by funding from American Heart Association, NIH and the University of Pennsylvania Research Foundation.

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